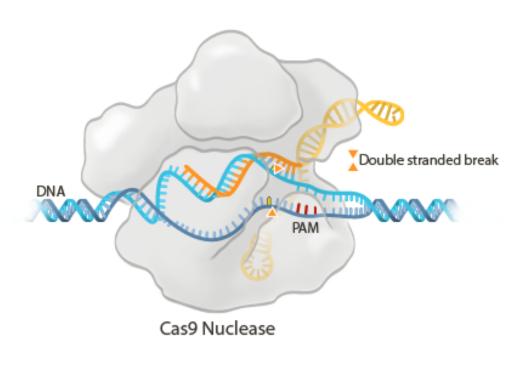
# editas

Directed evolution platforms and applications for engineering RNAguided nucleases

Barrett Steinberg, Derek Cerchione, Eric Tillotson, Joost Mostert, Chase Kelley, Jen DaSilva, Fred Harbinski, Hari Jayaram, Vic Myer

#### **O** Directed evolution to improve gene editing

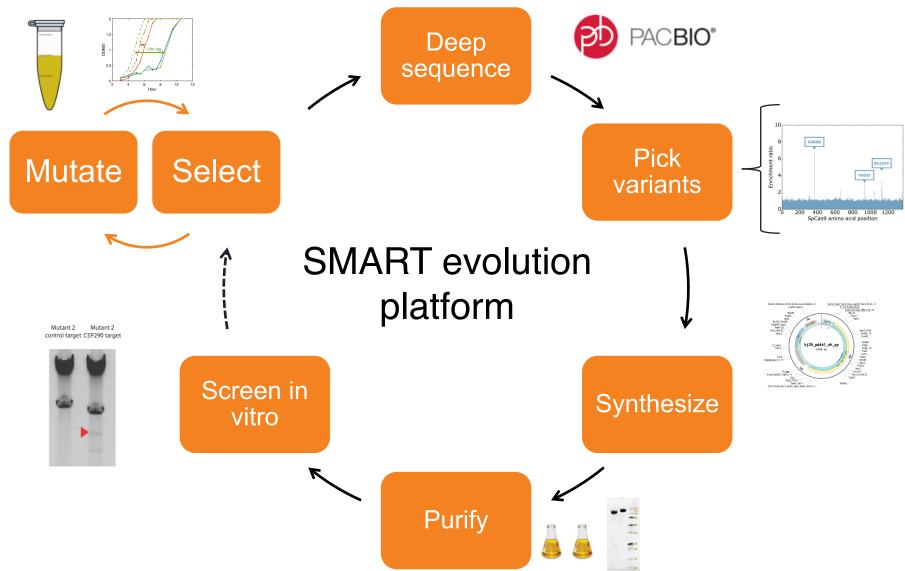
- CRISPR is an amazing tool with great potential in the clinic, but there are improvements to make
- Rational design is often difficult
- Design can be slow as more gene editing tools are discovered
- We are developing a directed evolution platform for quickly and effectively engineering gene editing tools



## O Building new systems for directed evolution of nucleases

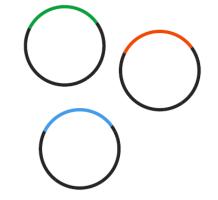
SMART libraries	<ul> <li>Improving mutagenesis over whole proteins</li> </ul>
Phage-based selection	<ul> <li>Competitive and fast evolution in liquid culture</li> </ul>
Specificity evolution	<ul> <li>Application to designing against identified off-targets</li> </ul>
PAM evolution	<ul> <li>Application to changing PAM preferences of enzymes</li> </ul>

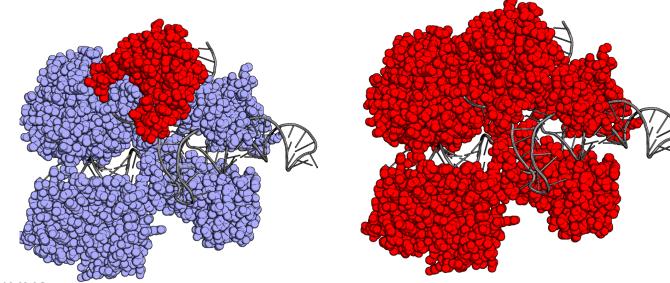
#### **O** Directed evolution pipeline



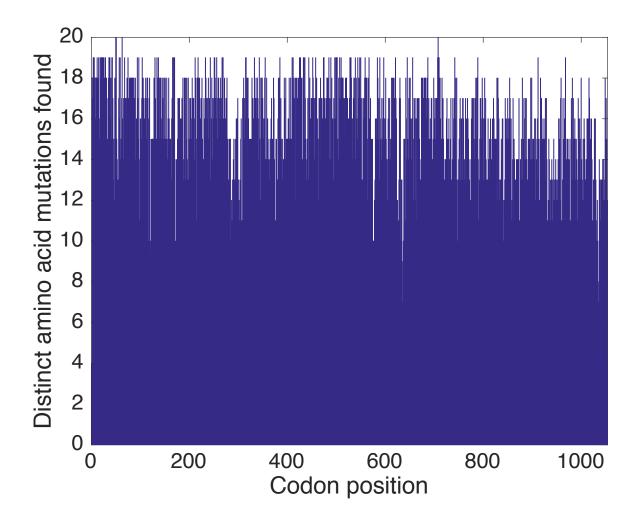
#### **Mutagenesis: SMART library creation**

- Scanning mutagenesis at random targets
- Codon mutations along entire cas9 gene
- Can be targeted to specific regions
- One-pot ~6hr protocol on dsDNA template
- High yield



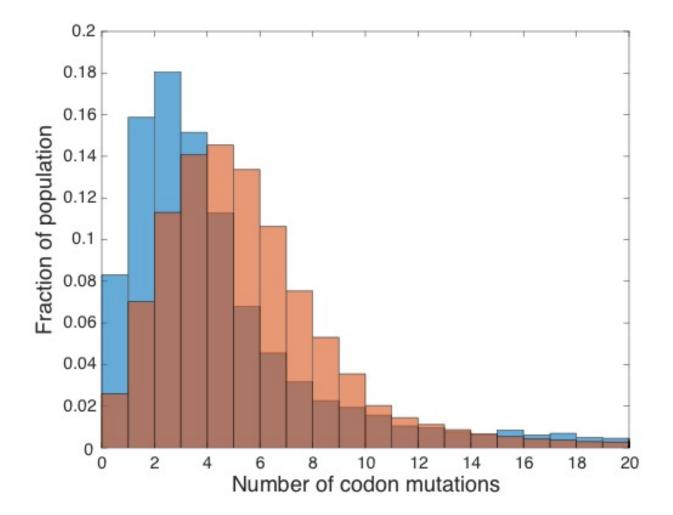


## SMART accesses all potential amino acid mutations throughout the protein



We can infer we are making every possible mutation, but is diversity of libraries overwhelming?

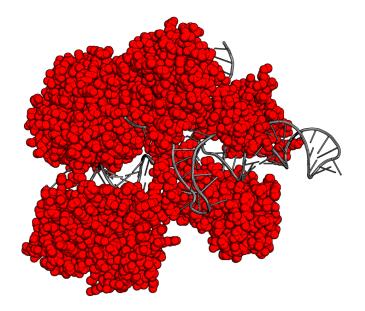
#### **SMART** mutation rate is tunable



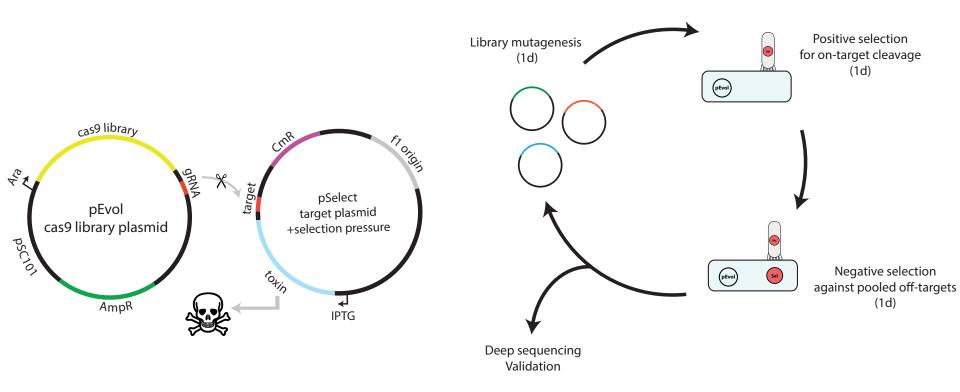
With a million transformants, we have 10X coverage of all single mutants and sample other mutations broadly © 2017 Editas Medicine

#### **CO** SMART libraries are comprehensive

- Comprehensive libraries containing every single amino acid mutation
- Not limited by region
- Using NNN codon replacement, but can create non-random mutations
- Better libraries lead to better hits

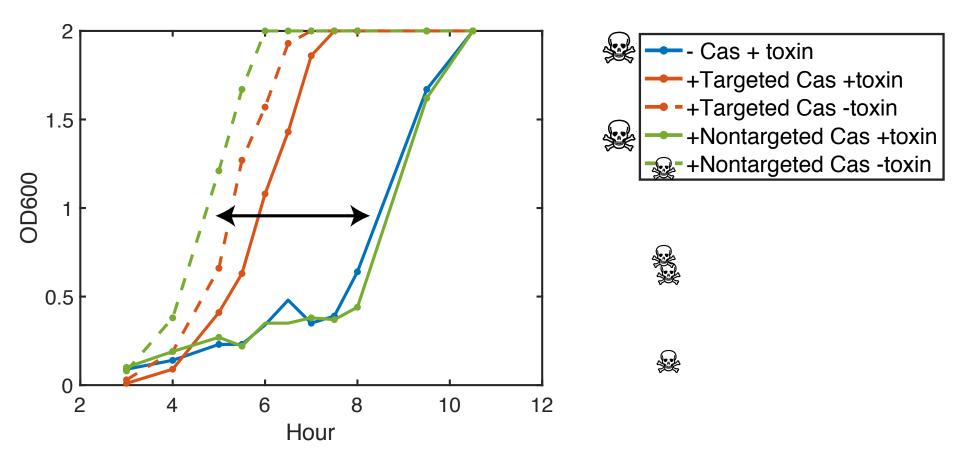


#### **O** Phage-based selection for or against cleavage

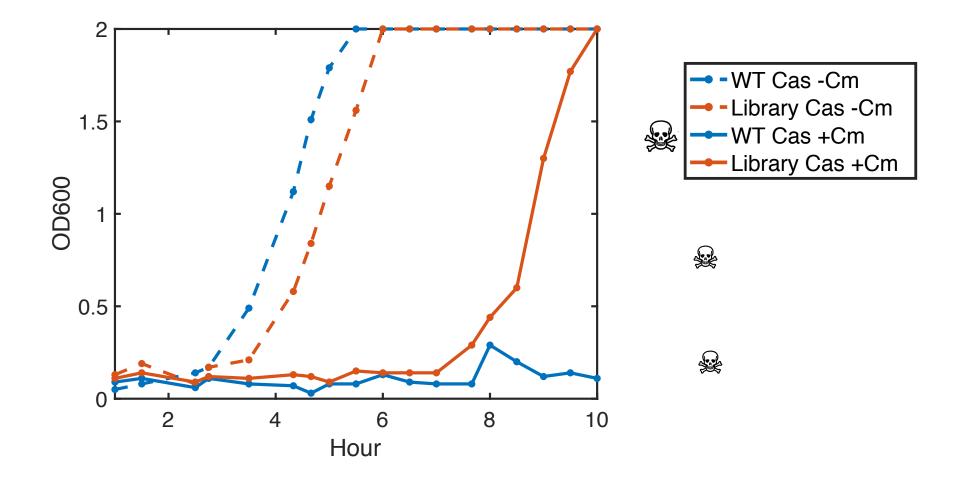




#### **O** Positive selection is effective in liquid culture

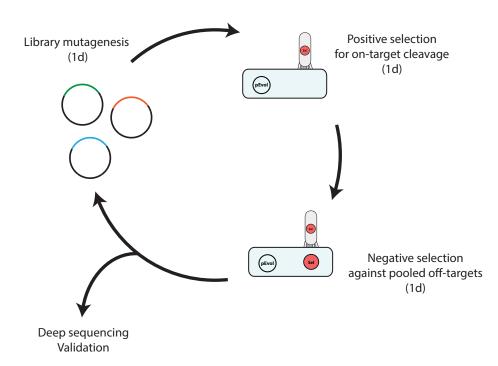


#### **Negative selection is effective in liquid culture**



#### **CO** Effective phage-based selection for nuclease activity

- Highly modular and fast
- Can handle large libraries
- Not a screen competition increases selective events
- Multiple selective conditions can be pooled together



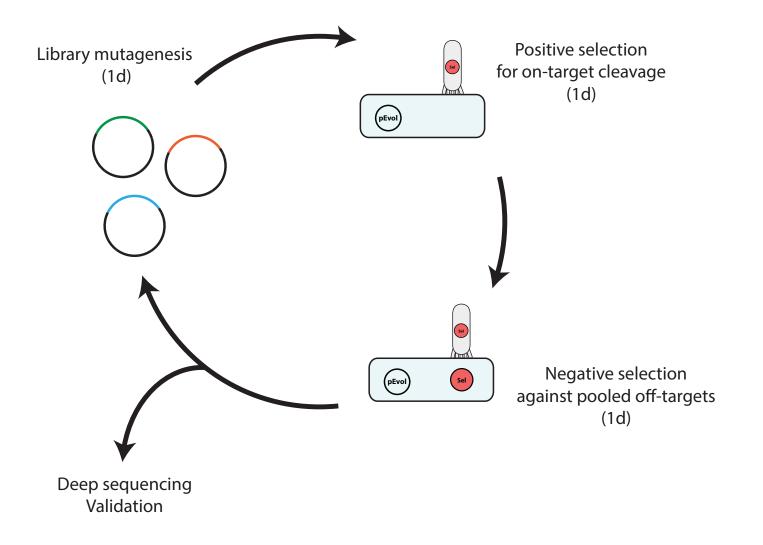
## O Building new systems for directed evolution of nucleases

SMART libraries	<ul> <li>Improving mutagenesis over whole proteins</li> </ul>
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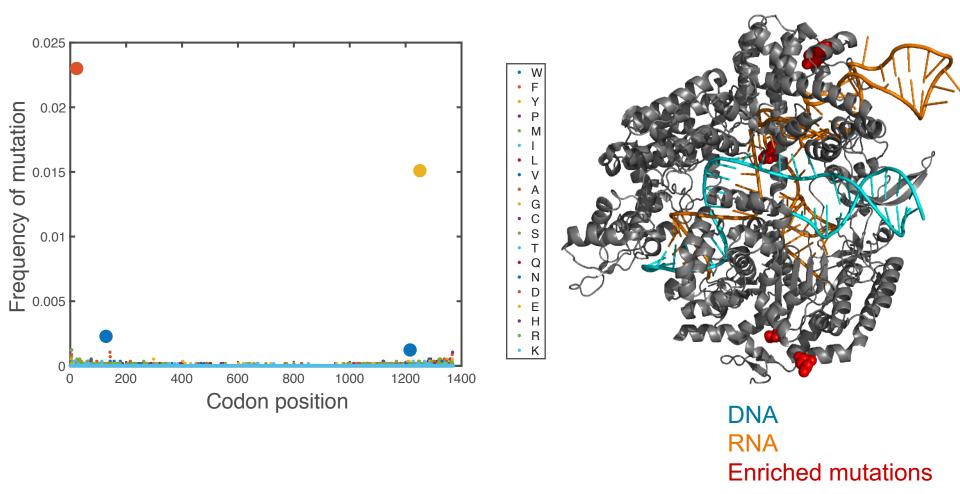
#### O Application: Selection against identified off-targets in Sp. Cas9

- Four off-targets were identified in edited T-cells
- We wanted to decrease off-target cutting while maintaining on-target efficacy
- Approach should be enzyme- and target-agnostic and tailorable to a specific guide
- Does not require structural information

#### **Selection against identified off-targets with phage**

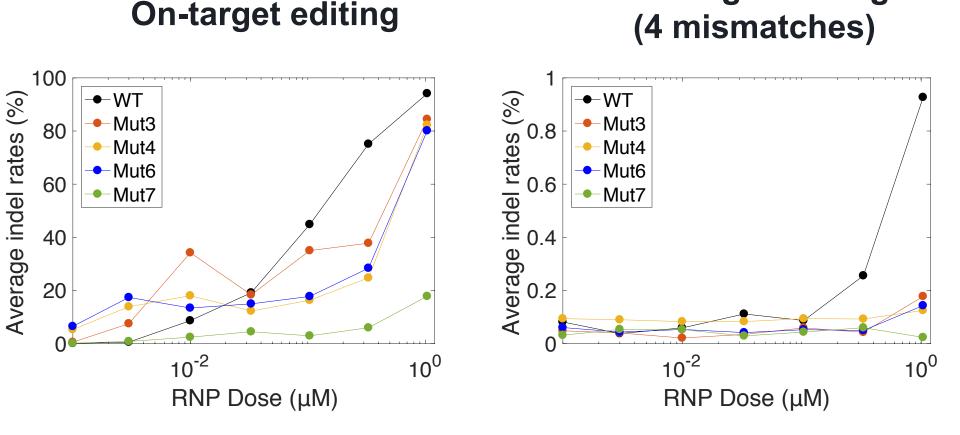


## Identified enriched cas9 mutations via deep sequencing



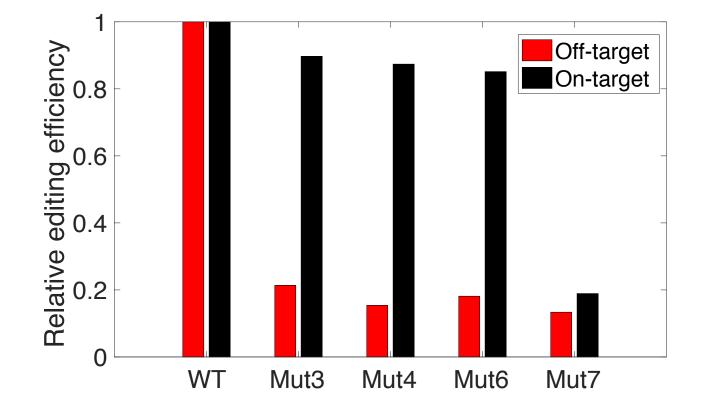
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Mutant proteins show decreased off-target editing and maintain on-target efficacy in T-cells

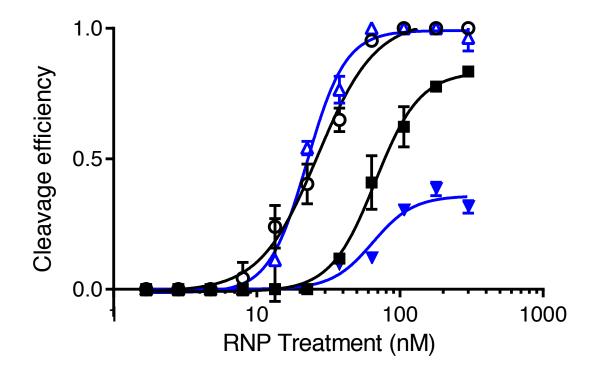


**Off-target editing** 

#### Mutant proteins show decreased off-target editing and maintain on-target efficacy in T-cells



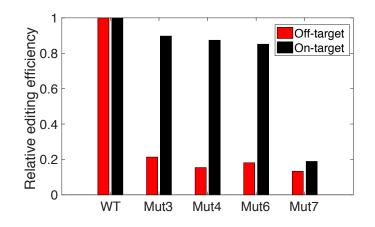
#### CO Combinations of enriched mutations reduce offtarget cutting in vitro



- wt OnTarget
- wt Off Target
- ▲ Mut6 On target
- ➡ Mut6 Off target

#### **CO** Evolution against identified off-targets

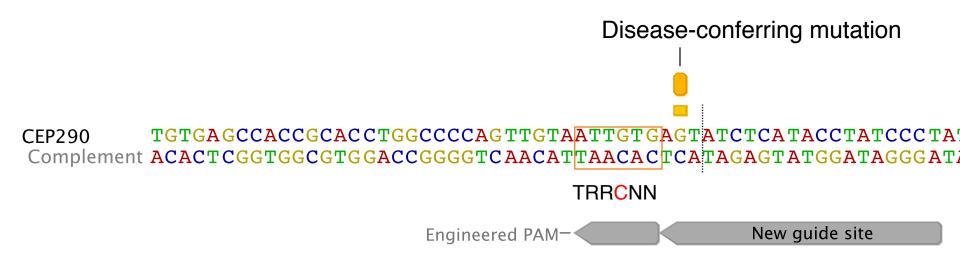
- We can evolve cas9 variants to mitigate identified off-targets
- Multiple off-targets can be selected against in each round
- We maintain on-target cleavage efficiency
- In vitro tests confirm lower max cutting may indicate competitive inhibition



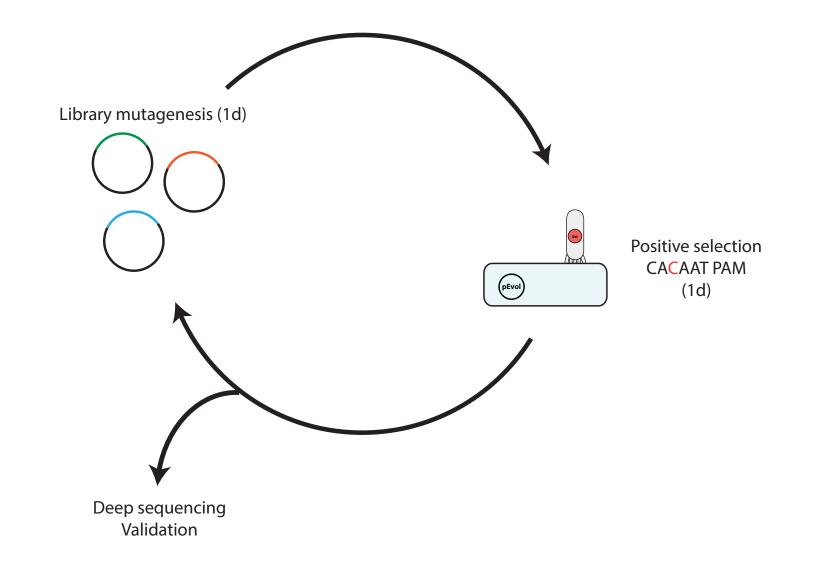
## **Evolution for alternate PAMs in Sa. cas9**

Wildtype Sa. Cas9 requires NNGRRT adjacent motif to cleave

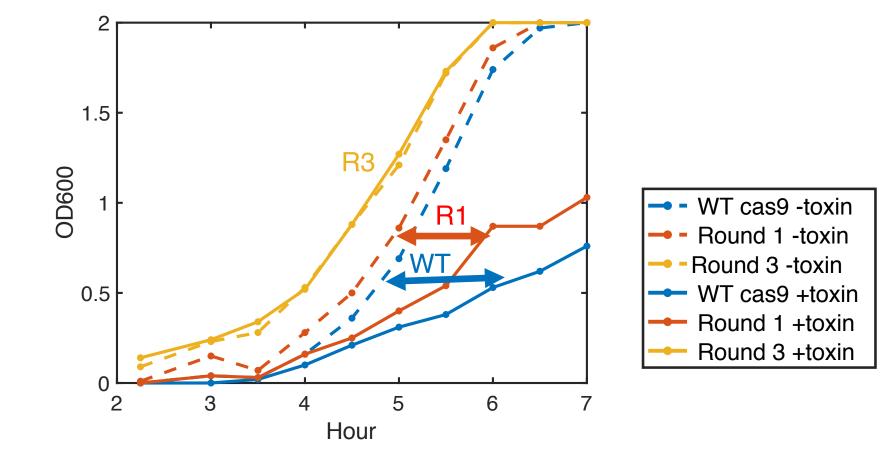
We wanted to evolve a change to NNCRRT to target a specific cut



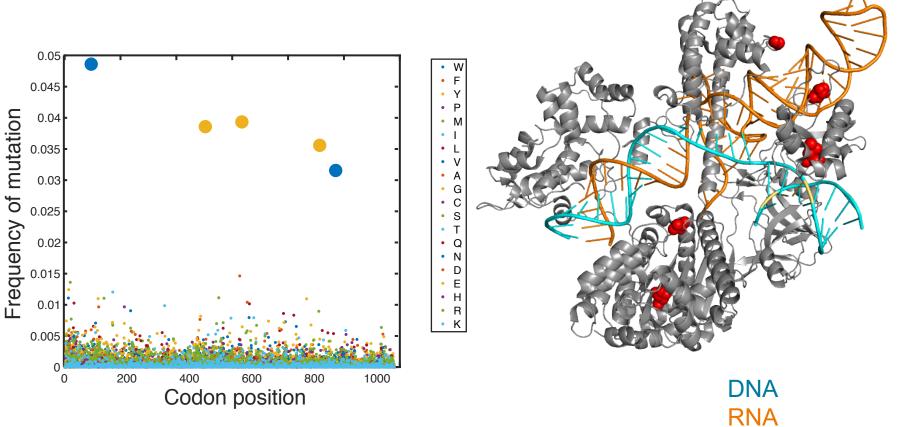
#### O Application: Evolution for alternate PAMs in Sa. cas9



#### **CO** Libraries have increasing fitness during evolution



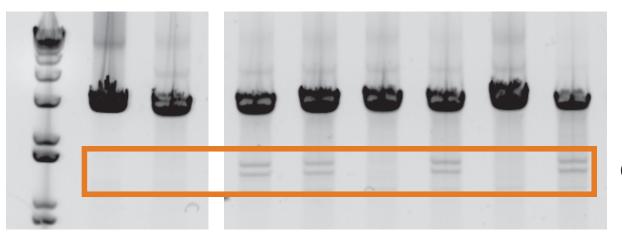
#### **CO** Identified hits via deep sequencing



Enriched mutations Altered PAM base

### CO Combinations of enriched mutants are functional for novel PAM cleavage

#### Neg WT Mut1 Mut2 Mut3 Mut4 Mut5 Mut6

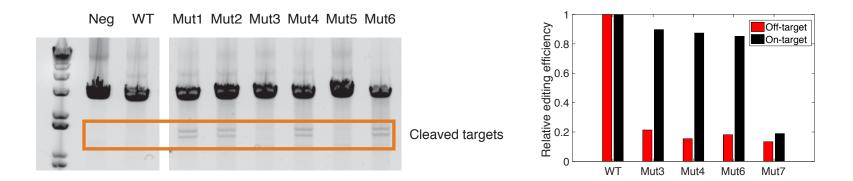


**Cleaved targets** 

We can evolve towards new sequence dependencies



- SMART mutagenesis creates highly diverse libraries
- Phage can be used in competitive evolution of nuclease activity
- We can selectively dial-down off-target activity while retaining on-target efficacy
- We can engineer new PAM-changing enzymes





- Derek Cerchione
- Hari Jayaram
- Editas NGS team
- Grant Welstead
- Morgan Maeder
- Vic Myer
- Editas Platform group







#### **CO** SMART libraries have low bias

