

In Vivo Proof of Concept for EDIT-102: A CRISPR/Cas9-Based Experimental Medicine for USH2A-Related Inherited Retinal Degeneration Caused by Mutations in Exon 13

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Usher Syndrome Type 2A and EDIT-102

- Mutations in the *USH2A* gene are a major cause of autosomal recessive retinitis pigmentosa (RP) and Usher syndrome Type II (USH2A). Patients with USH2A mutation have early hearing loss, progressive peripheral vision loss, and eventual legal blindness.
- USH2A* gene encodes the transmembrane protein Usherin which is localized in connecting cilia of photoreceptors and believed to play a role in stabilizing the photoreceptor outer segment
- c.2299delG mutation in exon 13 is the most common mutation in *USH2A* gene, causing a frameshift and truncated protein, which leads to ciliary defect. It is a common cause of inherited retinal degeneration (IRD).
- There are currently no approved disease-modifying therapies for RP caused by USH2A mutations
- We have developed EDIT-102, an experimental medicine for USH2A-related IRD through CRISPR/Cas9-mediated excision of USH2A exon 13.
- EDIT-102 is an AAV5 vector expressing *S. aureus* CRISPR/Cas9 driven by the photoreceptor-specific G protein-coupled receptor kinase 1 (GRK1) promoter and two U6 promoter-driven gRNAs that flank hUSH2A exon 13 (Figure 1).

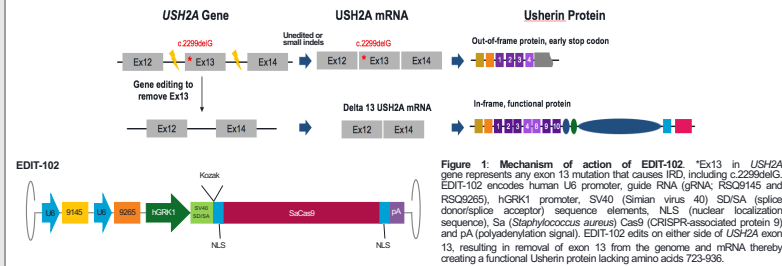


Figure 1: Mechanism of action of EDIT-102. Exon 13 in *USH2A* gene represents any exon 13 mutation that causes IRD, including c.2299delG. EDIT-102 encodes human U6 promoter, guide RNA (gRNA; RSG945 and RSG285), hGRK1 promoter, SV40 (Simian virus 40) SDSA (splice donor/splice acceptor) sequence elements, NLS (nuclear localization sequence), Sa (*Staphylococcus aureus*) Cas9 (CRISPR-associated protein 9) and pA (polyadenylation signal). EDIT-102 edits on either side of *USH2A* exon 13, resulting in removal of exon 13 from the genome and mRNA thereby creating a functional Usherin protein lacking amino acids 723-936.

USH2A Humanized Knock-In (KI) Mouse Models

- A humanized *USH2A* KI mouse model (hUSH2A^{2299delG}) was generated, where mouse *Ush2a* exon 12 was replaced with the human *USH2A* exon 13 containing the 2299delG mutation and a portion of the flanking introns (Figure 2A).

- Around 17% of *USH2A* transcripts in heterozygous hUSH2A^{2299delG} mouse retinal tissue contain human exon 13, as measured by Nanostring.

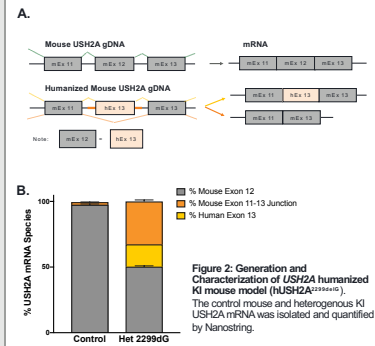


Figure 2: Generation and Characterization of USH2A humanized KI mouse model (hUSH2A^{2299delG}). The control mouse and heterozygous KI USH2A mRNA was isolated and quantified by Nanostring.

Methods

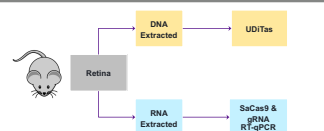


Figure 3: Experimental Schematic for EDIT-102 Pharmacology Studies. Heterozygous hUSH2A^{2299delG} KI mice were subretinally injected with EDIT-102 or vehicle. At different time points, retinas were harvested and split equally for DNA & RNA assays. On-target *USH2A* gene editing was determined using the retinal DNA in a unidirectional targeted sequencing (UDI-Tas) method. Expression levels of SaCas9 and guide RNA (gRNA) were evaluated using the retinal RNA in a real time quantitative polymerase chain reaction (RT-qPCR) assay.

Productive USH2A Gene Editing in EDIT-102 treated Mice

- Subretinal injection of 1 µl of AAV5 vector AAV5-GKR1-GFP show 30% transduction of mouse neural retina in previous study (Maeder 2019).
- The total on-target edits is calculated by including large deletions, inversions, AAV/plasmid integrations and small indels.
- Inversions and deletions were previously defined as productive edits that generate *USH2A*^{Δ13} mRNA. Normalized productive editing is obtained by multiplying the percentage of productive editing with the transduction multiplier, as shown below:

$$\text{Transduction area of retina: } 30\% \\ \text{Transduction multiplier: } 100/30 = 3.3 \\ \text{Normalized Productive Editing} = \text{Productive Editing} * 3.3$$

Results

Normalized Productive Editing of *USH2A* Gene and Cas9 mRNA/gRNA Expression Increased with the Increase of EDIT-102 dose

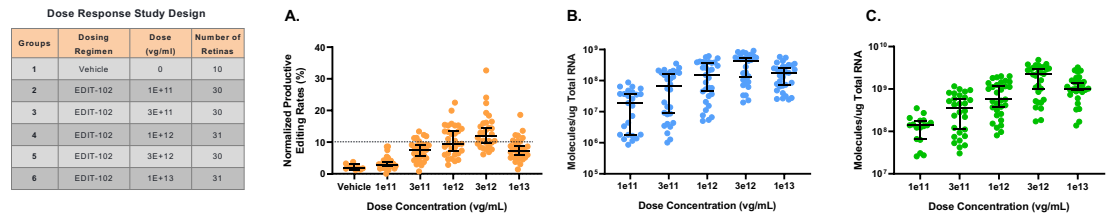


Figure 4: Dose Response Study For EDIT-102. The figure shows the normalized productive editing rates (Median ± 95% confidence interval) (A), Cas9 mRNA (B) and gRNA (C) expression levels in hUSH2A^{2299delG} KI mice. Each dot represents a single retinal sample and the dotted line represents 10% normalized productive editing of photoreceptors.

The level of normalized productive editing in the mouse retina increases with the dose of EDIT-102 from 1e11 to 3e12 vg/mL and decreases at 1e13 vg/mL. We expect a therapeutic benefit when 10% foveal cone photoreceptors are productively edited (Geller 1992, Geller 1993). The dose response of EDIT-102 from 3e11 to 3e12 vg/mL achieve 10% or greater productive editing in the treated mouse retinas.

Normalized Productive *USH2A* Gene Editing and Cas9 mRNA/gRNA Increased with Time

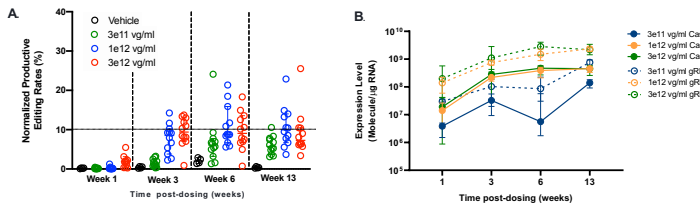


Figure 5: Time course study for EDIT-102. The figure indicates the normalized productive editing of *USH2A* gene (A) and Cas9 mRNA and gRNA expression (B) over time. The Vehicle group includes 16 retinas and the dose 3e11, 1e12 and 3e12 vg/ml of EDIT-102 groups have 47, 48 and 48 retinas. Each point represents an individual mouse eye harvested at various timepoints from 1 to 13 weeks post-dosing.

The median level of normalized productive editing and expression level of Cas9 mRNA and gRNA, at the doses of 1e12 and 3e12 vg/ml, plateaued at 6 weeks and was maintained at a stable level over 13 weeks post-dose.

Total Editing Correlates with gRNA and Cas9 mRNA Expression

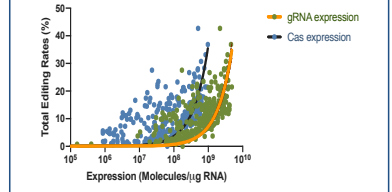


Figure 6: Total *USH2A* Gene Editing Rates and Cas9 mRNA and gRNA Levels in EDIT-102 Treated hUSH2A^{2299delG} KI Mice. Each data point indicates a single mouse retina processed in EDIT-102 dose-response and time-course studies, 1, 3, 6, or 13-week post subretinal injection. Curve fitting between total non-normalized editing and Cas or gRNA expression was fitted using a non-linear regression model using GraphPad Prism. Spearman rank correlation coefficient with P-value <0.0001.

Conclusions

- Pharmacokinetic and pharmacodynamic studies indicated Cas9 and gRNA expression correlated with total editing. Dose response of EDIT-102 from 1e12 to 3e12 vg/ml achieved close to 10% or greater normalized productive editing in the mouse retinas.
- All animals used in the pharmacological studies demonstrated tolerability as determined by clinical observations and body weight measurement.
- These data support the continued development of EDIT-102 for the potential treatment of USH2A-related IRD caused by mutations in exon 13.

References

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