CRISPR-Cas12a Gene Editing Enhances Functional Metabolism of Natural Killer Cells and Enables Tumor Cell Cytolysis in Metabolically Stressful Conditions That Inhibit Effector Cell Function

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OBJECTIVE

To evaluate the function of healthy donor-derived CRISPR-Cas12a-engineered natural killer (NK) cells in the nutrient-deprived and metabolically unfavorable tumor microenvironment as a proxy for CRISPR-Cas12a-engineered NK cells derived from induced pluripotent stem cells (iPSCs).

RESULTS

INTRODUCTION

- The tumor microenvironment is nutrient-deprived, due to competition between effector and tumor cells for essential nutrients, and enriched in immunosuppressive metabolites (eg, lactic acid) due to Warburg Metabolism.
- Effective anti-tumor cell therapies must be able to function in the tumor microenvironment.
- NK cells derived from healthy donors or iPSCs (iNK cells) with CRISPR-Cas12a-mediated knockout of cytokine-inducible SH2-containing protein (CISH) and transforming growth factor beta receptor II (TGFBR2) genes have demonstrated resistance to TGF-β inhibition and increased tumor control.1-3

METHODS

- CRISPR-Cas12a-edited CISH-/-/TGFBR2-/- NK cells derived from healthy donors were used as a model for CRISPR-Cas12a-engineered iNK cells.
- NK cells were challenged with SK-OV-3 ovarian tumor spheroids + 10 ng/mL TGF- β in the unfavorable metabolic conditions shown in the figure below, in isolation or combinatorially in a multifactorial matrix, and assayed for tumor cytotoxicity and inflammatory cytokine production



The cytotoxicity of NK cells was also assayed with SK-OV-3 ovarian tumor spheroids that were selectively evolved to grow in nutrient-deprived and/or high lactate media.

The mitochondrial function and fitness of NK cells were assayed using the Seahorse Cell Mito Stress Kit







CONCLUSIONS

- CRISPR-Cas12a-edited CISH--/TGFBR2-- NK cells demonstrated superior cytotoxicity and enhanced metabolic function in metabolically unfavorable conditions compared with unedited controls.
- This nutrient-deprived and immunosuppressive metabolic tumor model will be used to evaluate the metabolic and cytotoxic functions of CRISPR-Cas12a-engineered iNK cells with edits targeting metabolic pathways.

- 1. Wong KK, et al. SITC Annual Meeting 2020: Abstract 145 2. Borges CM, et al. ASH Annual Meeting 2020:Abstract 1436
- 3. Moon J-I, et al. ASH Annual Meeting 2020: Abstract 3257

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