



Characterization of gRNAs and Ribonucleoproteins for CRISPR Applications

Steve Wolk

VP, Analytical Chemistry & Site Head, Boulder

Editas Medicine

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Steven Wolk is an employee and shareholder of Editas Medicine, Inc.



- Editas Analytical Chemistry Group
 - Jean-Noel Lemercier
 - Chrysa Latrick
 - Pranjali Ghude
 - Andrew Melie
 - Bridget Moffet
- Pallavi Gambhire (Analytical Development, Cambridge)
- Jared Clark, Ananya Dubey-Kelsoe (IonDX)
- Bruce Eaton (CBO, previously SVP of Chemistry)
- Keith Jarvis and the Editas Boulder PD Team



Introduction to CRISPR and RP-IP-UPLC Purity Methods

- Brief Introduction to CRISPR
 - gRNAs and RNPs

Characterization of Secondary Structure and Multimer Formation

- gRNAs
 - methods for characterizing intramolecular folding and multimer formation
 - secondary structure predictions, PAGE, SEC, Ion Mobility
- CRISPR enzyme complex (RNP)
 - impact of these structures on complex formation
 - PAGE, SEC, Ion Mobility, IEX

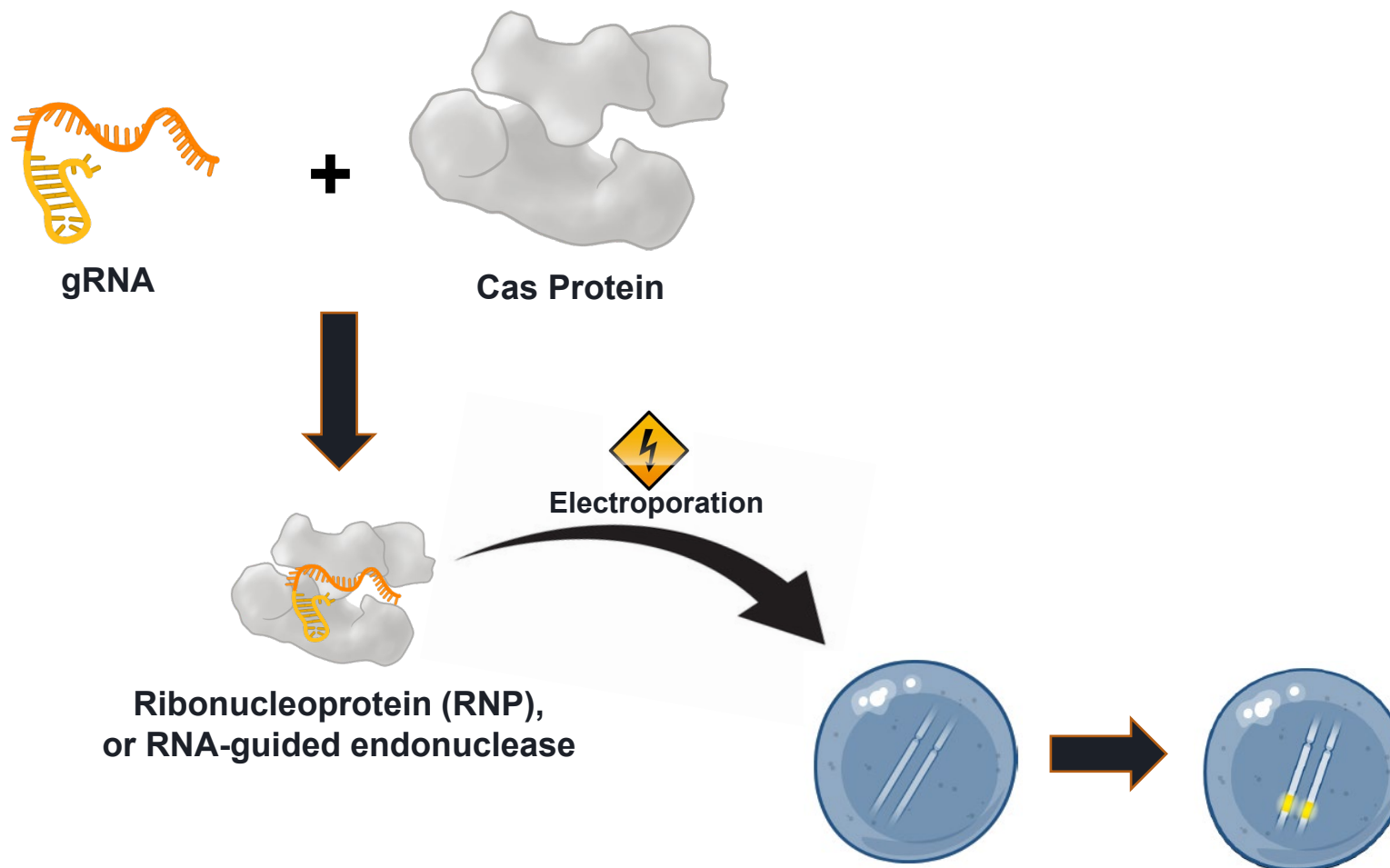
Concluding Remarks and Acknowledgements



CRISPR Basics

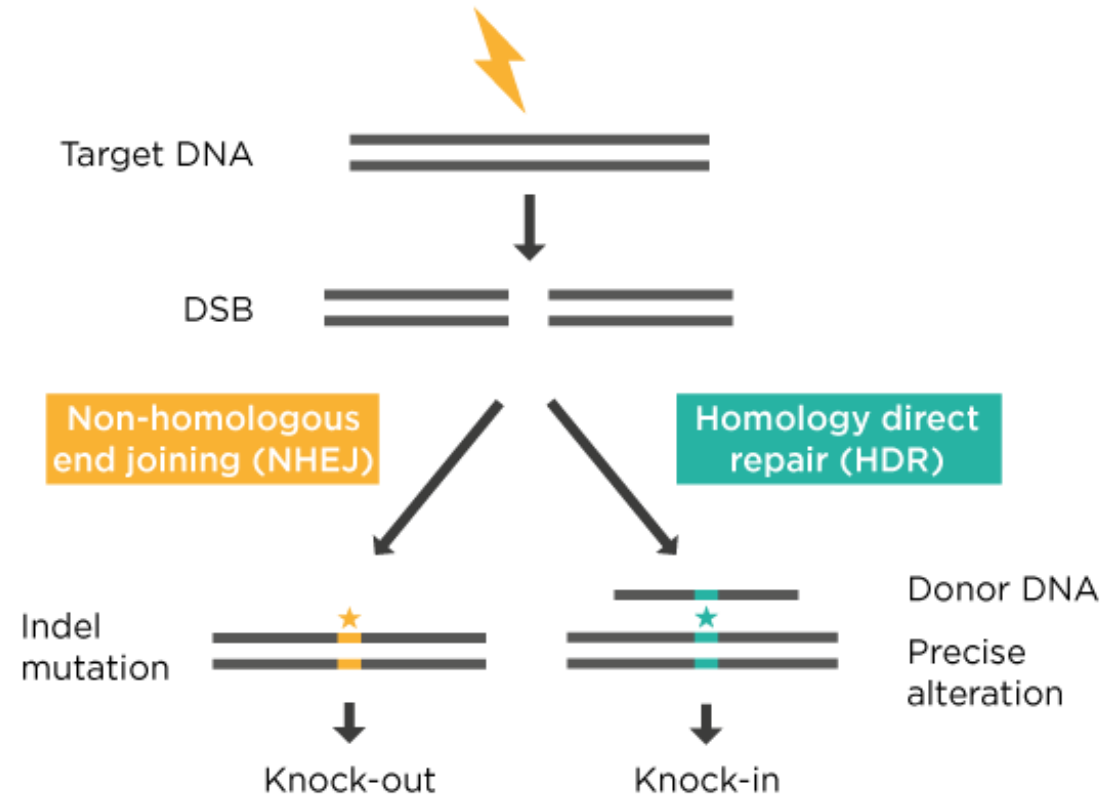


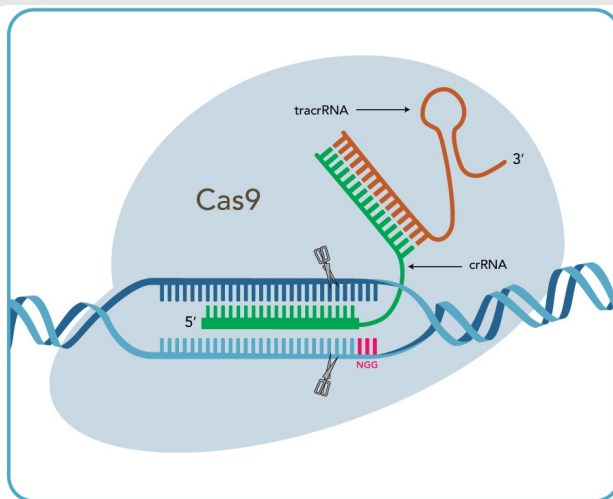
RNP: The Simple View of a Cell Based Medicine





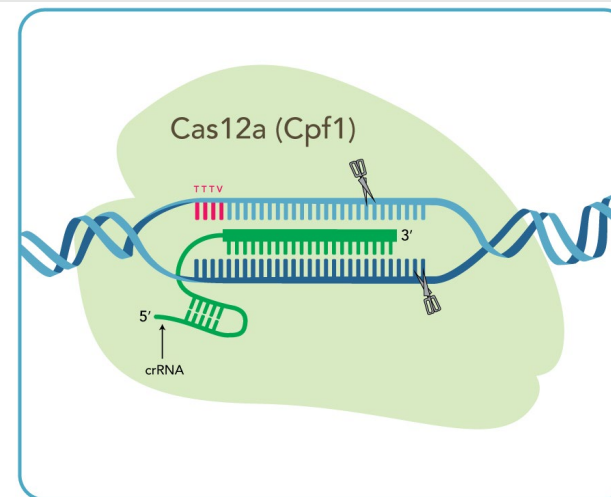
Knock-outs and Knock-ins





Cas9

- Separate **crRNA** and **tracrRNA** (can be synthesized as single guide RNA (sgRNA) (~100mer))
- G-rich PAM: **NGG**
- Blunt-ended cut
- Endonuclease domains:
 - RuvC-like (cleaves non-target strand)
 - HNH (cleaves target strand)
- Target sequence on 5'-terminus of the gRNA



Cas12a

- Single, shorter **crRNA** guide (40mer)
- T-rich PAM: **TTTN**
- staggered DNA cut with 4-5 nt overhangs
- Endonuclease domain: RuvC-like only
- Target sequence on 3'-terminus of gRNA
- **Shorter guides → higher purity, fewer competing structures**
- **3'-target region → better specificity**

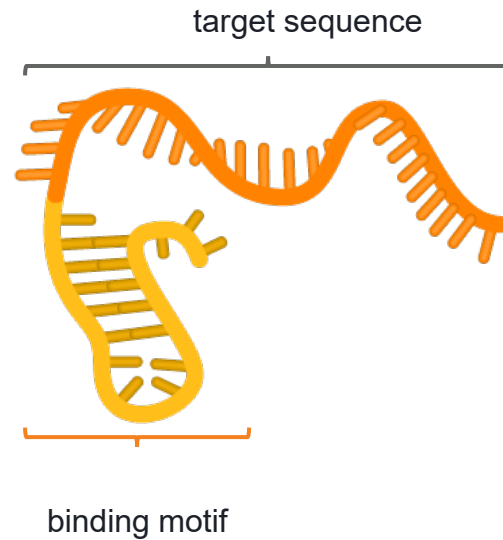


gRNA Characterization

It's Complicated.....



- This is how we like to think the gRNA looks:
- Which it does in the RNP crystal structures

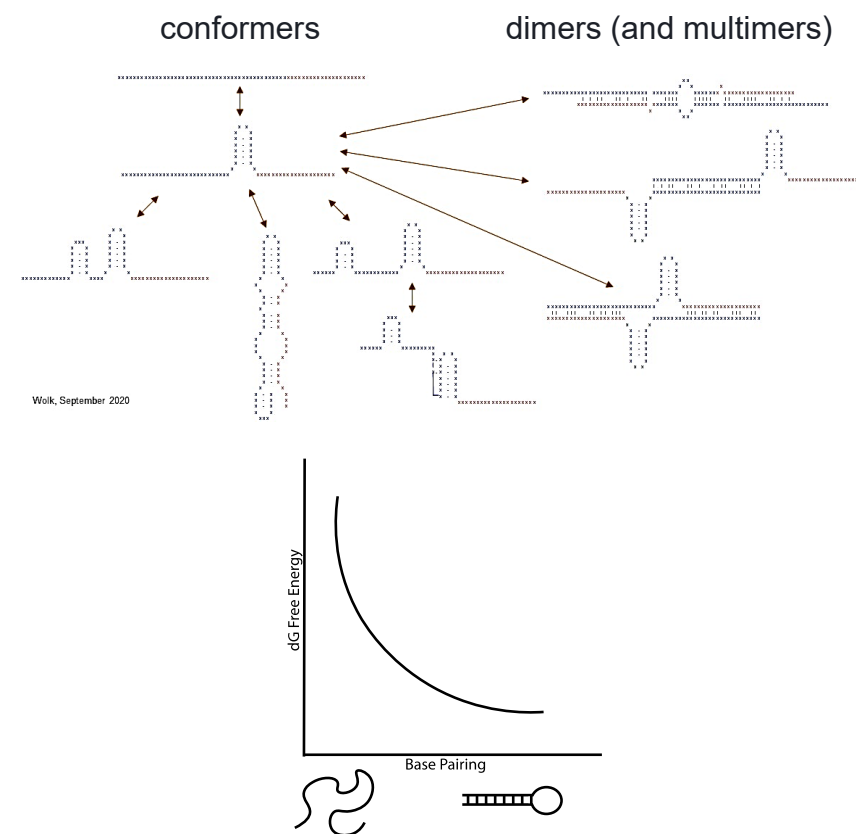




- This is how we like to think the gRNA looks:
- Which it does in the RNP crystal structures



- The situation is likely much more complicated
- Gets more complicated with increasing length



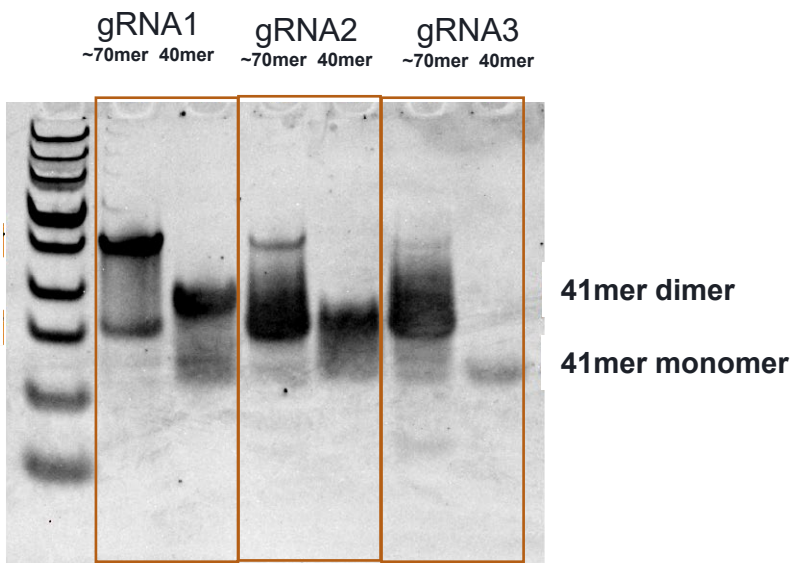


■ Guide Characterization

- (Purity/impurity Profile by LC/MS – not addressed today)
- Conformational Analysis
 - secondary structure predictions
 - PAGE
 - SEC
 - ion mobility



Dimerization for gRNA is Sequence Dependent



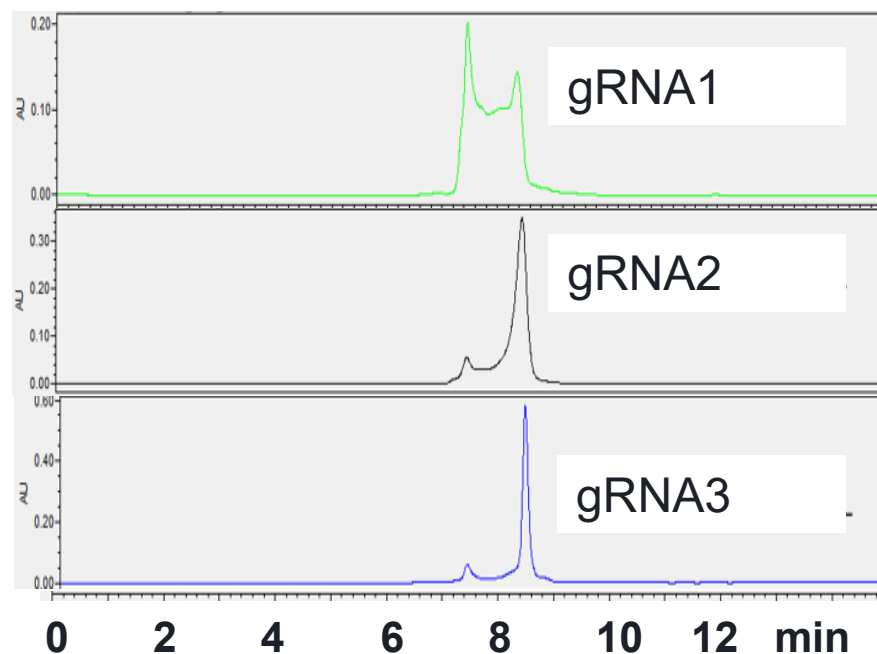
34 mM Tris, 66 mM HEPES, 75 mM NaCl, 2 mM MgCl₂, pH 7.5

guide	41mer	~70mer
gRNA1	dimer	dimer
gRNA2	dimer	monomer
gRNA3	monomer	monomer



Impact of gRNA Multimers

SEC of 3 gRNAs (~70mer versions): only change is the target sequence within the gRNA
(gRNAs analyzed on a Waters UPLC-SEC column)

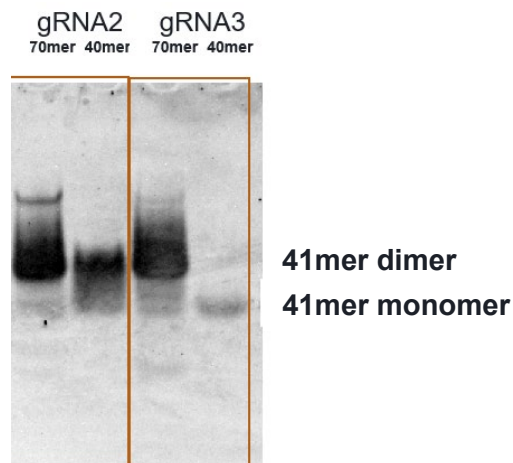


PAGE Results:

guide	41mer	~70mer
gRNA1	dimer	dimer
gRNA2	dimer	monomer
gRNA3	monomer	monomer

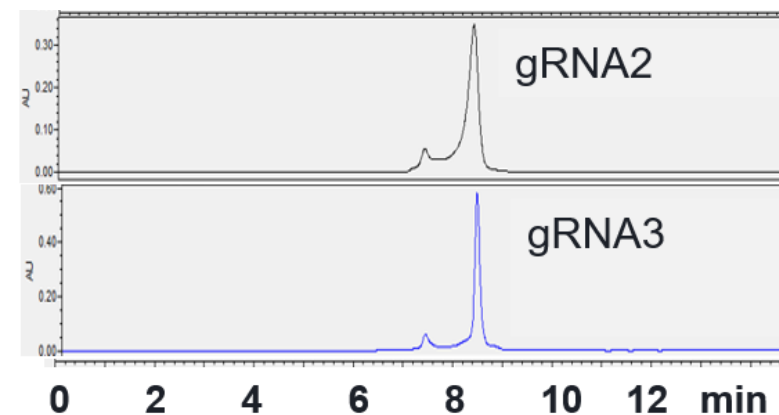


HOS for apo-gRNA is Sequence Dependent



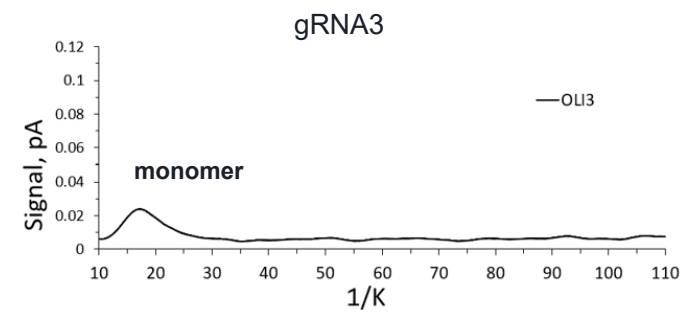
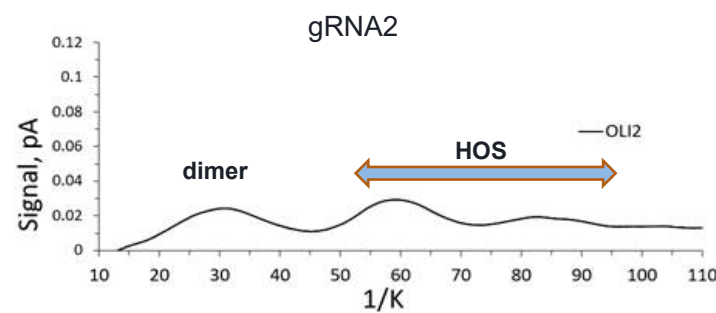
34 mM Tris, 66 mM HEPES, 75 mM NaCl, 2 mM MgCl₂, pH 7.5

SEC (~70mers)



150 mM NaCl, 100 Na-Phos, pH 7.4

Ion Mobility Data (~70mers), 5 mM (NH₄)OAc, pH 7.4
IonDX IMgenius



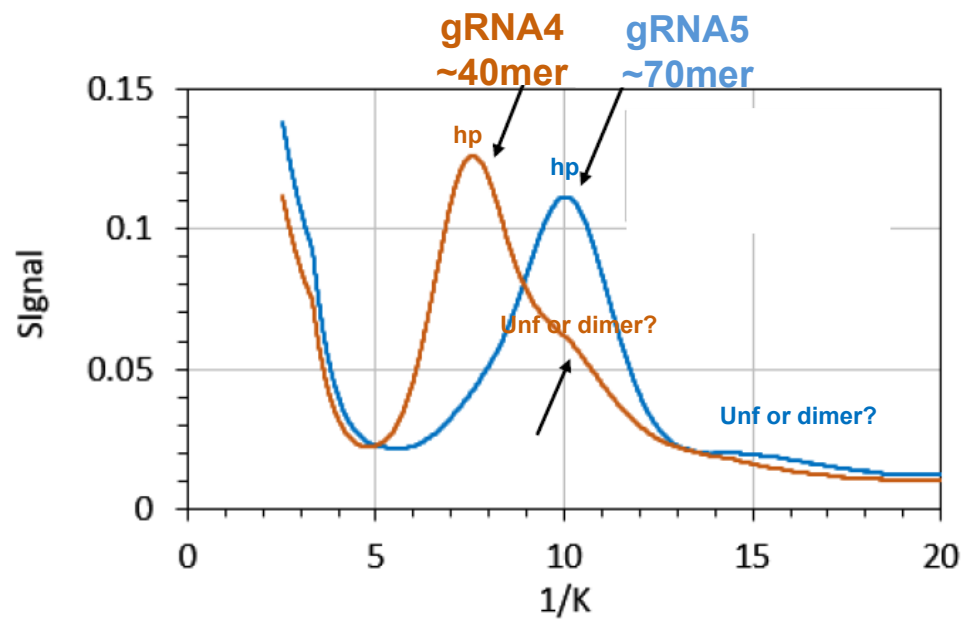
$1/K \propto$ collisional cross-section (size)



Detection by IM

Two additional gRNAs (40mer and ~70mer)

- 25 mM $(\text{NH}_4)_2\text{CO}_3$, pH 8.5
- earlier IM hardware parameters (resulting in different $1/K$ values)





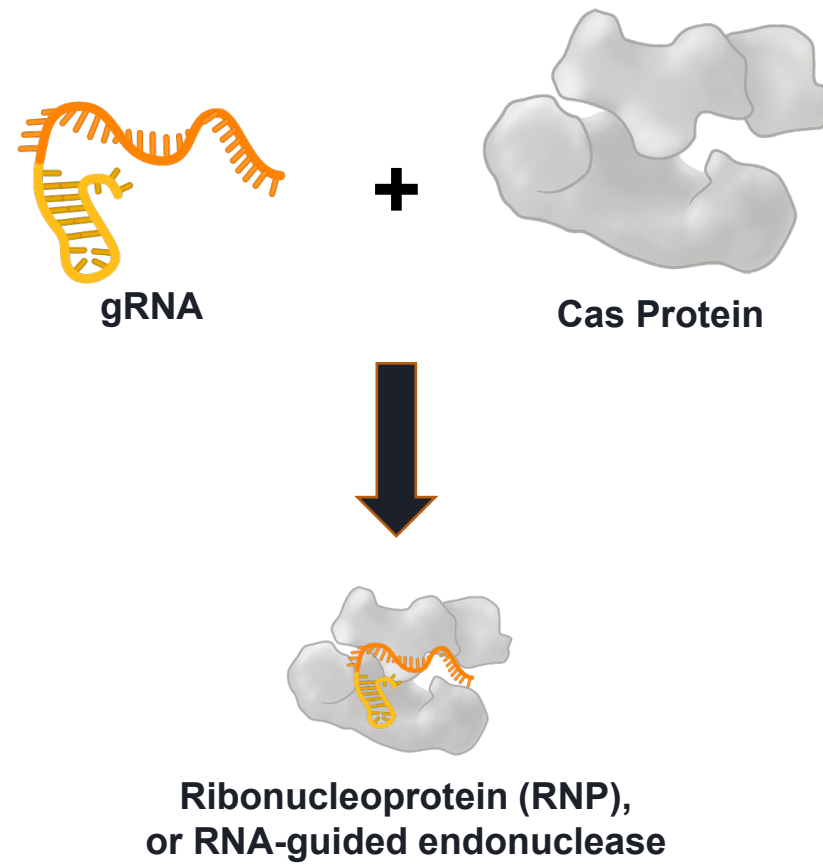
guide	Secondary structure prediction	PAGE	SEC	IM
	Vienna RNA-fold	75 mM NaCl 34 mM Tris 66 mM HEPES 2 mM MgCl ₂ pH 7.5	150 mM NaCl 100 Na-phos pH 7.4	5 mM (NH ₄)OAc pH 7.4
gRNA2	prone to multimer formation	mostly monomer	mostly monomer (exchange observed)	all HOS
gRNA3	not prone to multimer formation	monomer	mostly monomer	monomer

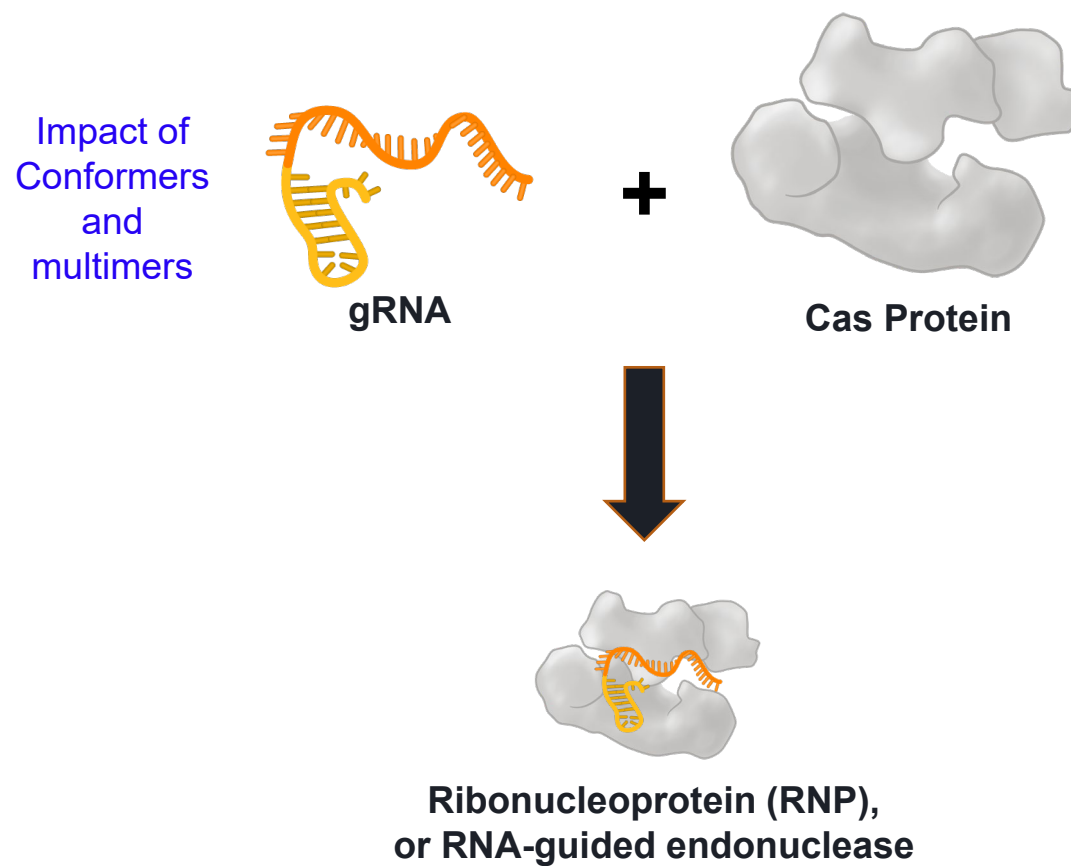
RNP Characterization

It's More Complicated.....



RNP: The Simple View





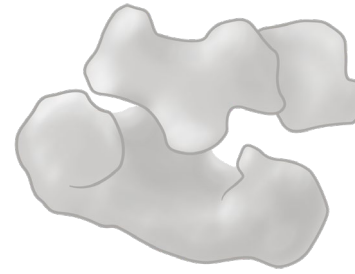


RNP: The Less, Less Simple View

Impact of
Conformers
and
multimers

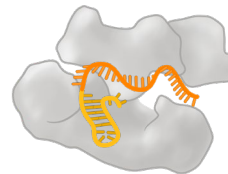


+



Cas Protein

- truncates
- PTMs
- disulfide variants
- aggregates (soluble/insoluble)
- degradants (oxi, deamidation)



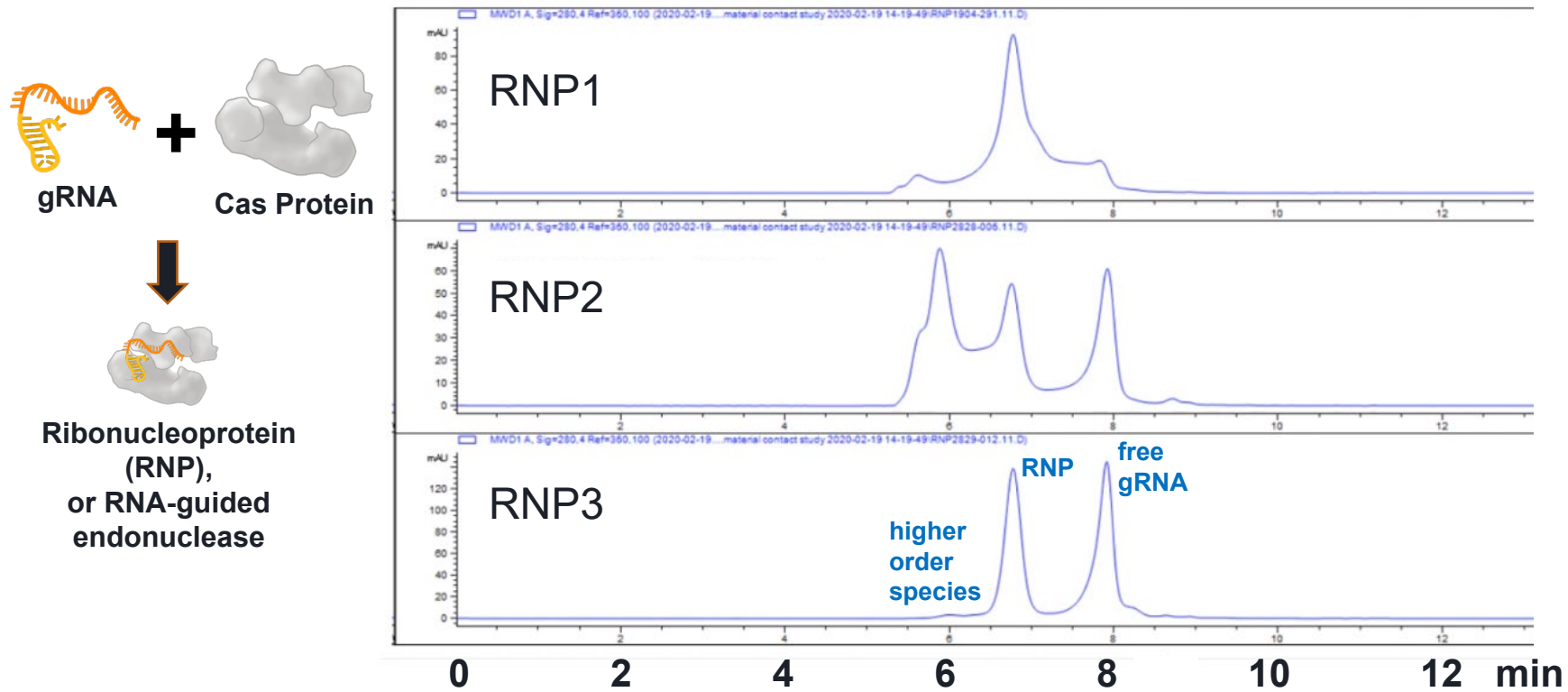
**Ribonucleoprotein (RNP),
or RNA-guided endonuclease**

- multiple stoichiometries
- aggregates (soluble/insoluble)
- dynamics/exchange



Impact of gRNA Multimers

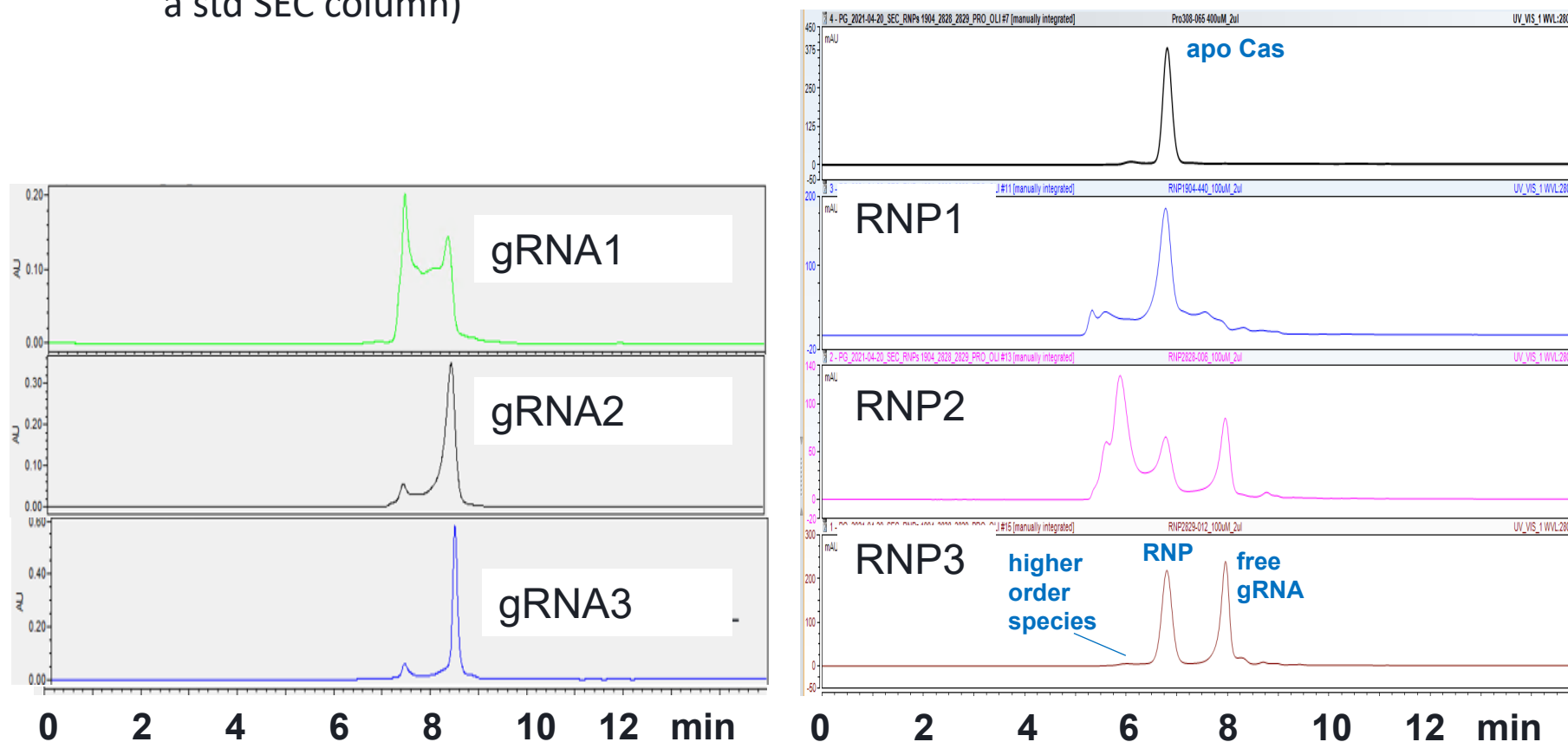
SEC of 3 RNPs: only change is the target sequence within the gRNA





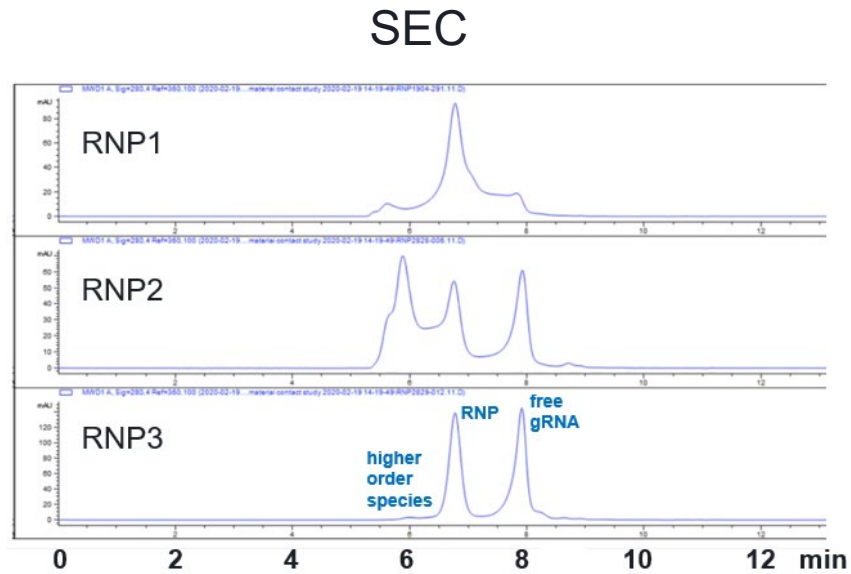
Impact of gRNA Multimers

SEC of 3 RNPs: only change is the target sequence within the gRNA
(gRNAs analyzed on a Waters UPLC-SEC column; RNPs analyzed on a std SEC column)

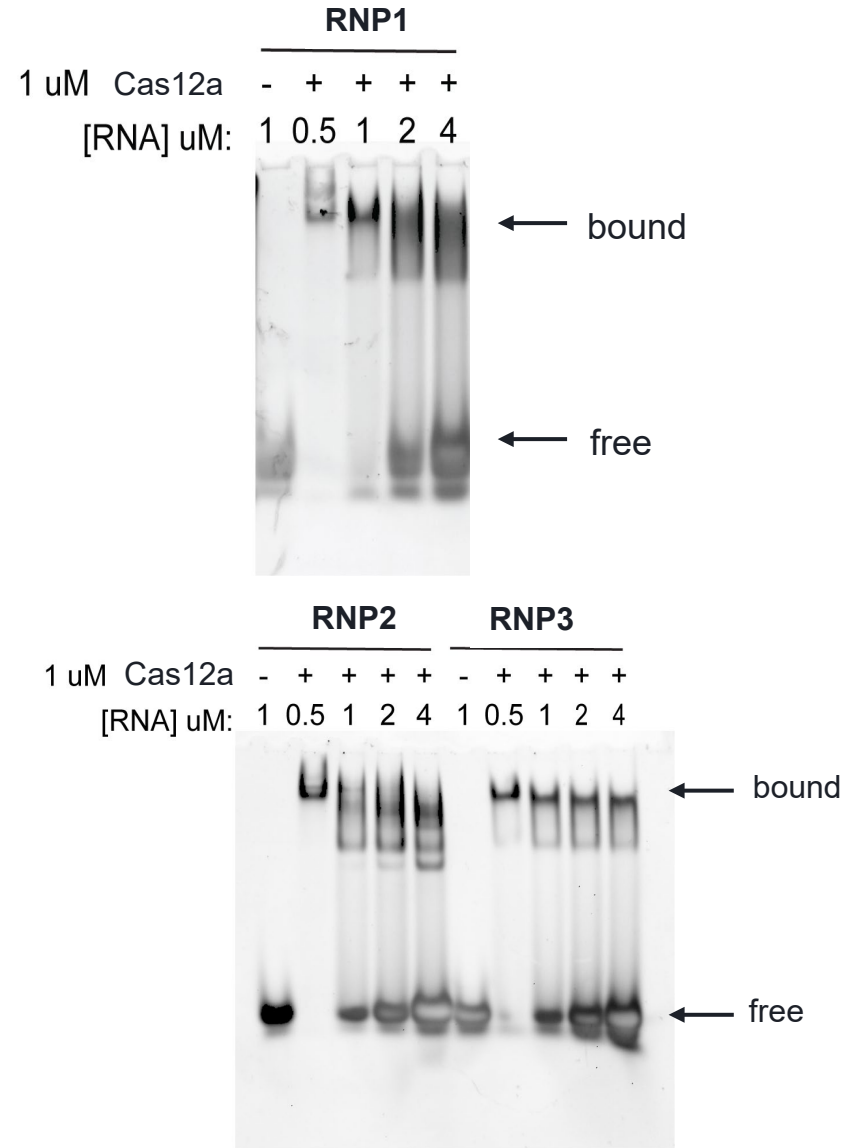




SEC and Gel Shift Data Correlate

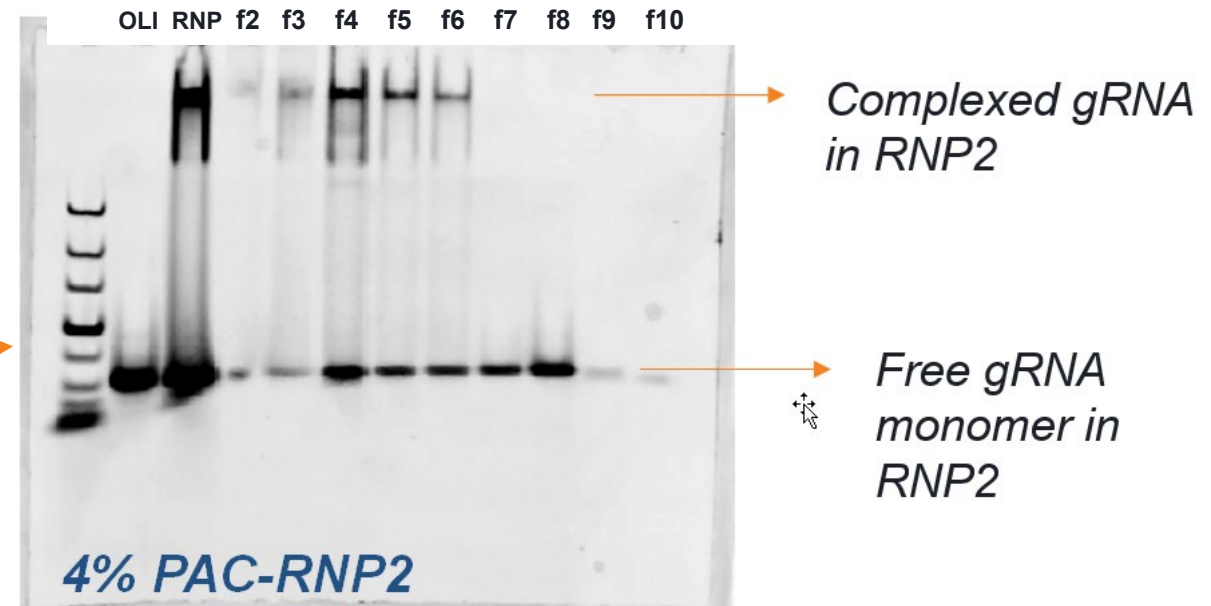
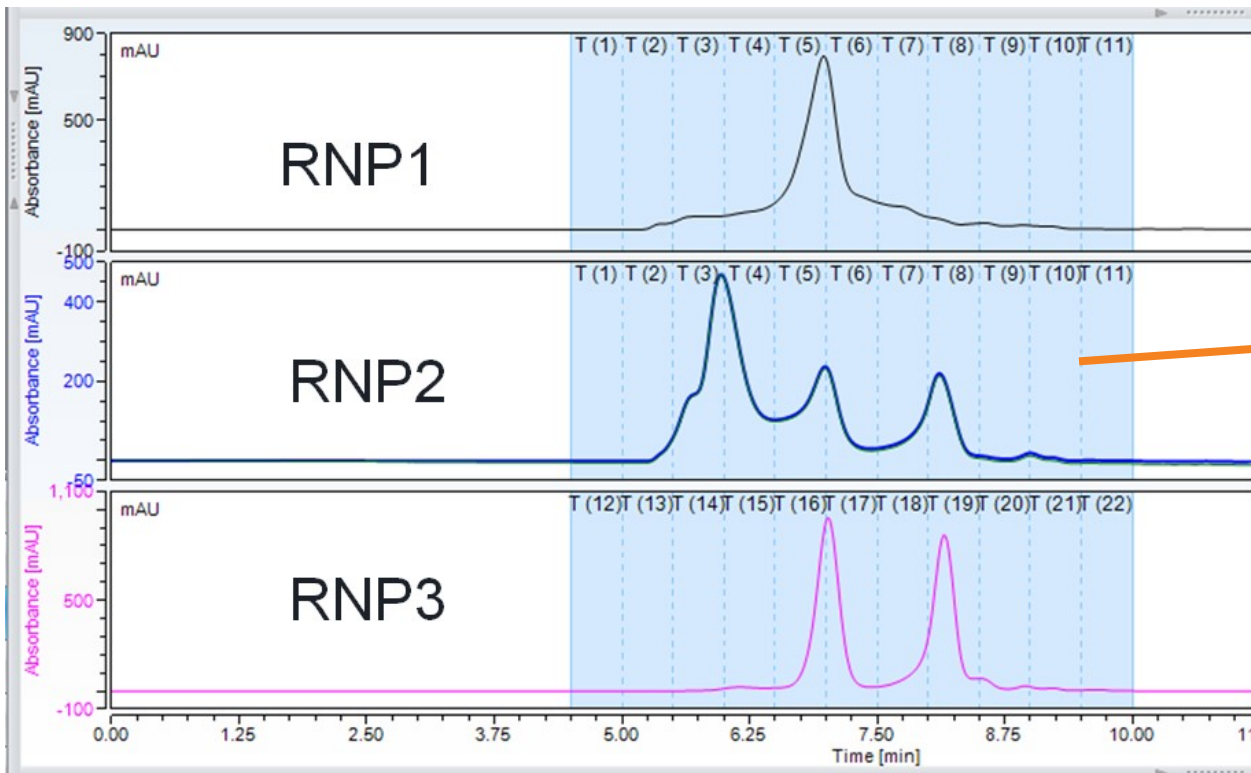


SEC analysis of 2:1 gRNA:PRO

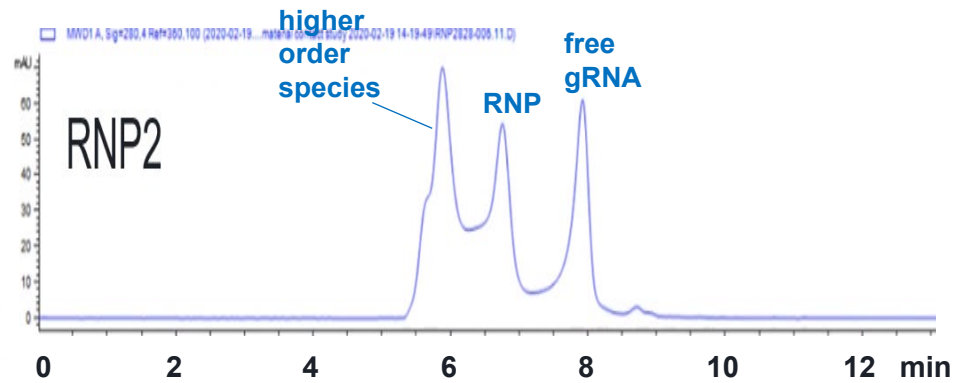




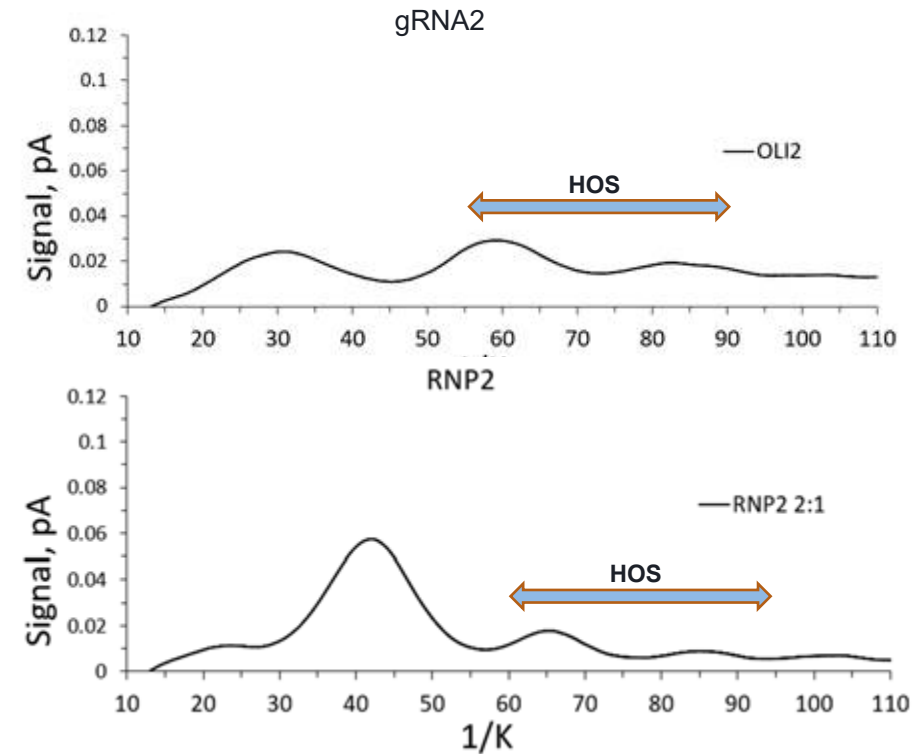
See Poster T.1 (Ghude et al.)



- Lane 1: LMW DNA ladder
- Lane 2: gRNA2 (unfractionated)
- Lane 3: RNP2 (unfractionated)
- Lane 4-12: SEC-Fractions of RNP2 (F2-F10)



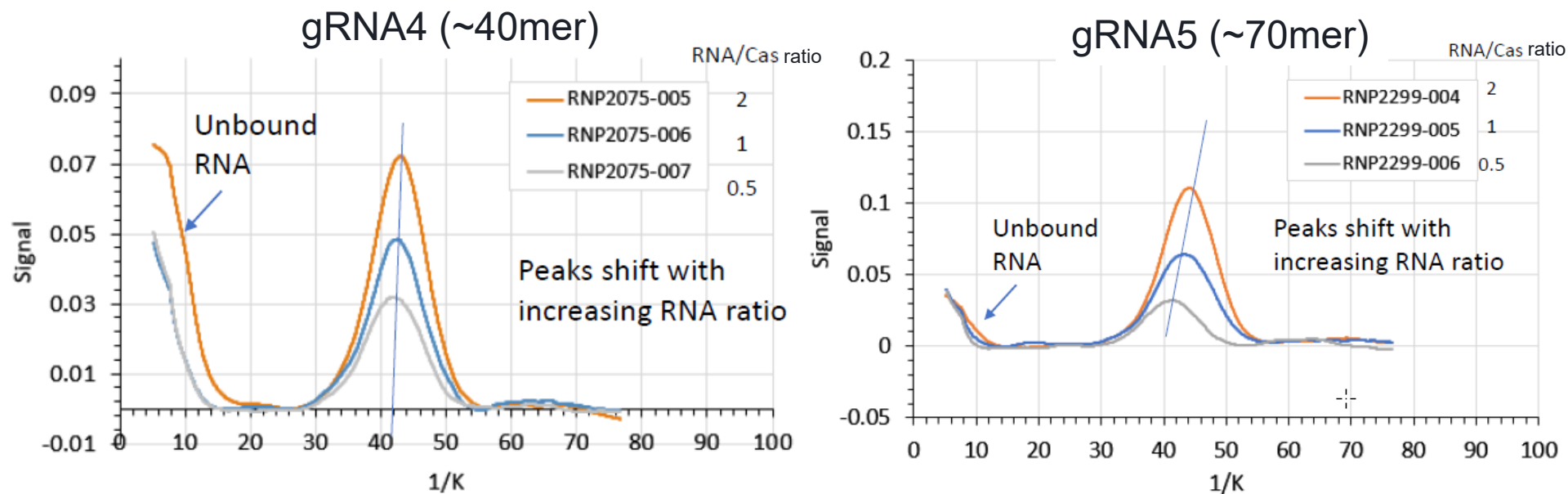
IM data in 5 mM NH_4OAc , pH 7.4



1/K	MW _{calc}
41.97	159 kDa
65.08	332 kDa
85.15	519 kDa



RNPs in 25 mM $(\text{NH}_4)_2\text{CO}_3$, pH 8.5

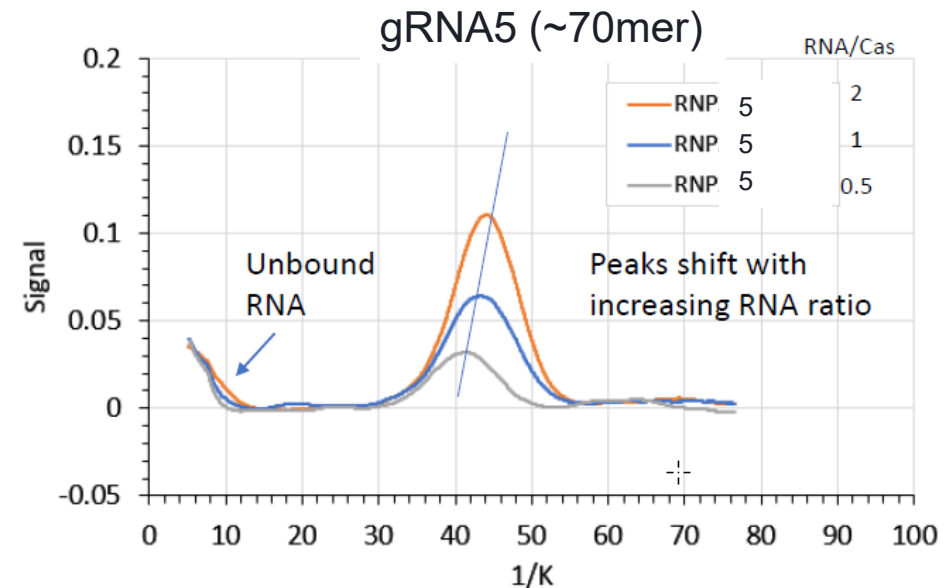
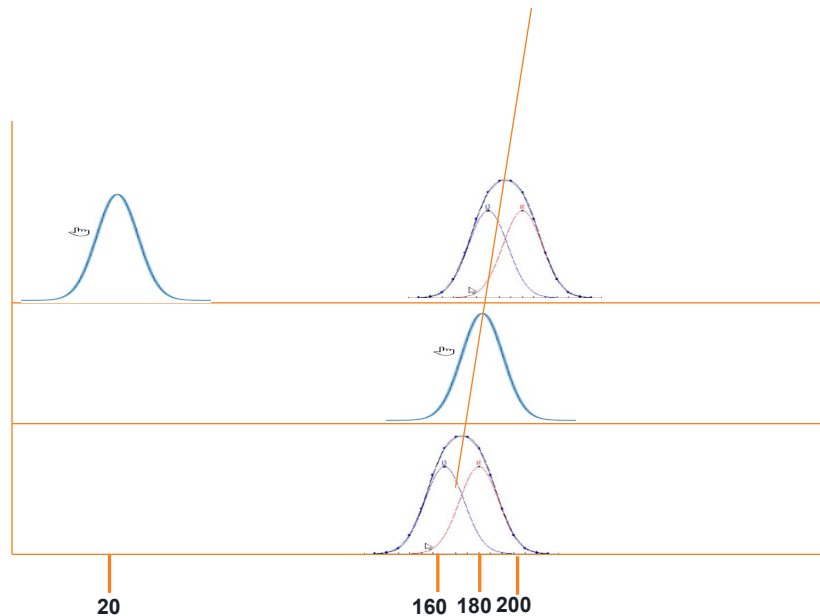




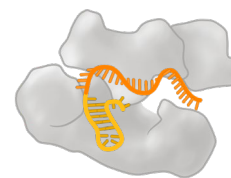
RNPs via IM: Varying the gRNA:Cas12a ratio

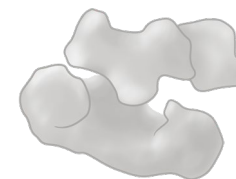


- Likely interpretation of the slope in the observed $1/K$:
 - 0.5:1 = superposition of two slightly resolved peaks
 - apo protein (160 kDa) and 1:1 (180 kDa))
 - 1:1 = mostly 1:1
 - 2:1 = superposition of two slightly resolved peaks
 - 1:1 (180 kDa) and 2:1(200 kDa) + apo-guide
 - $\Delta(1/K) < \text{peak width}$







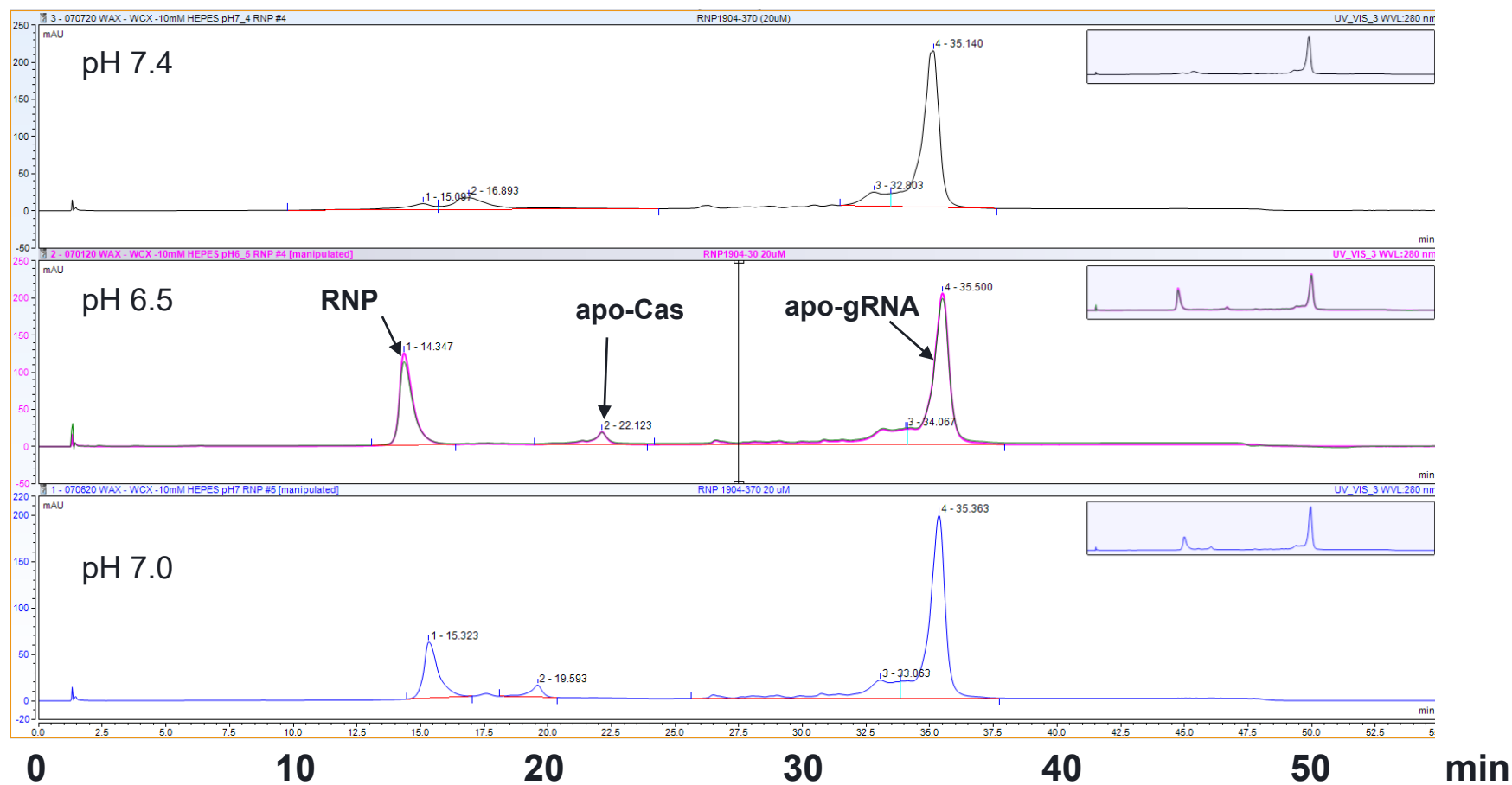


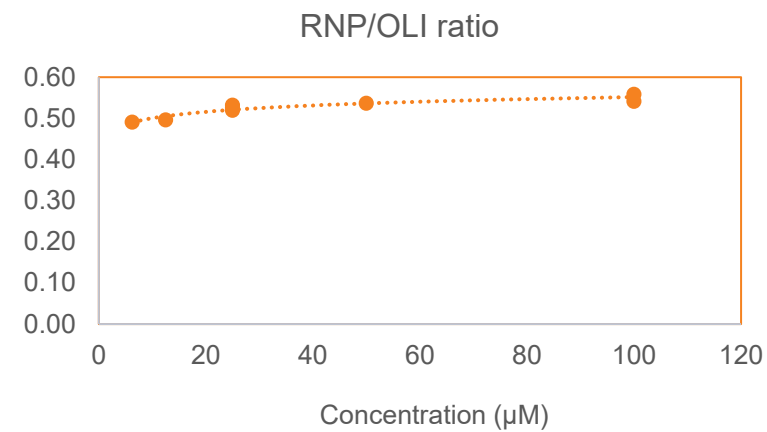
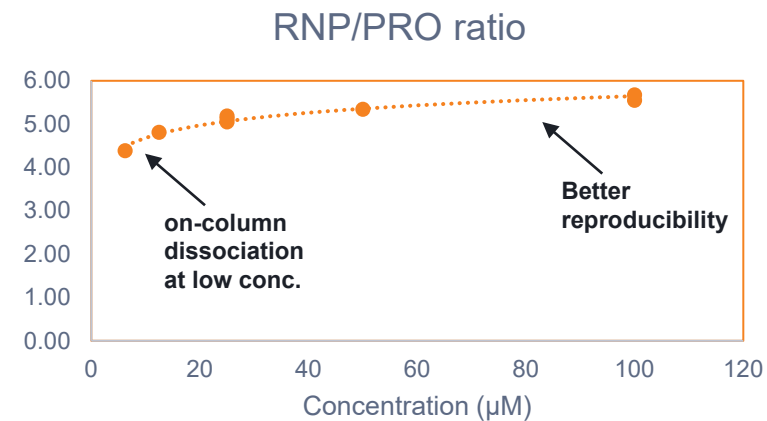
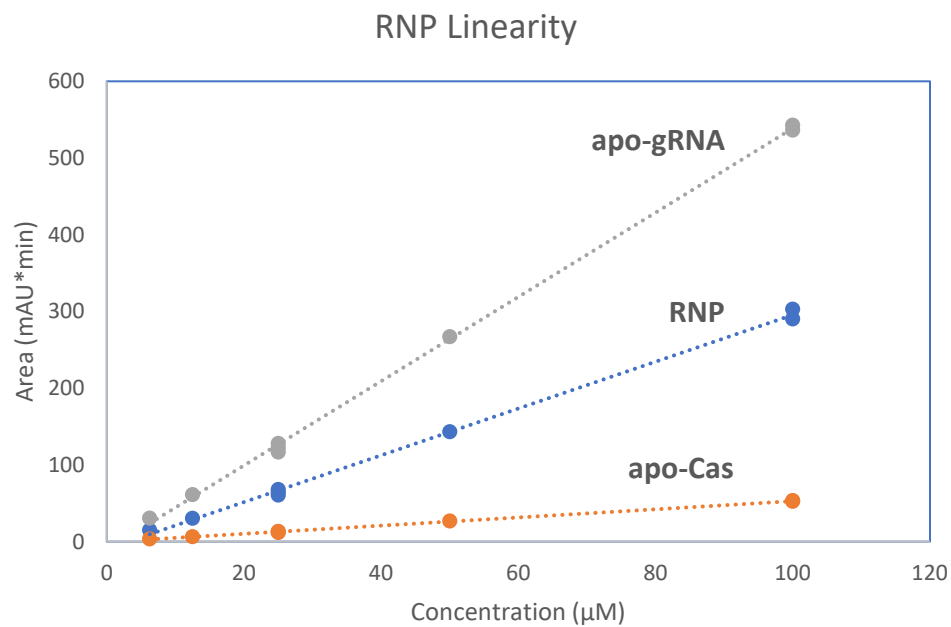
++

mode	gRNA	RNP	Cas
WCX	void	in between	retain
WAX	retain	in between	void
WCX-WAX	retain	retain	retain



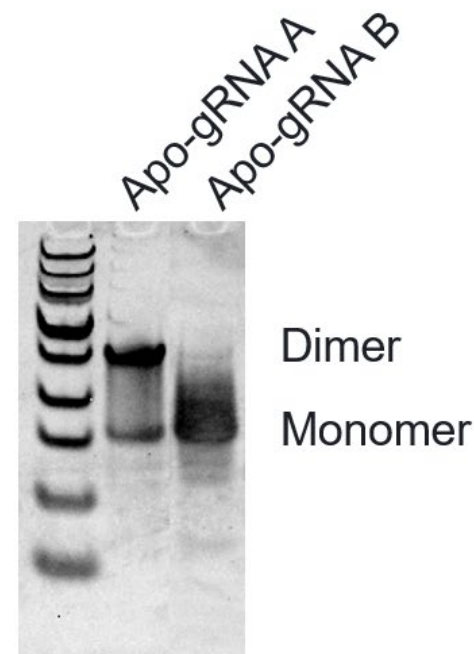
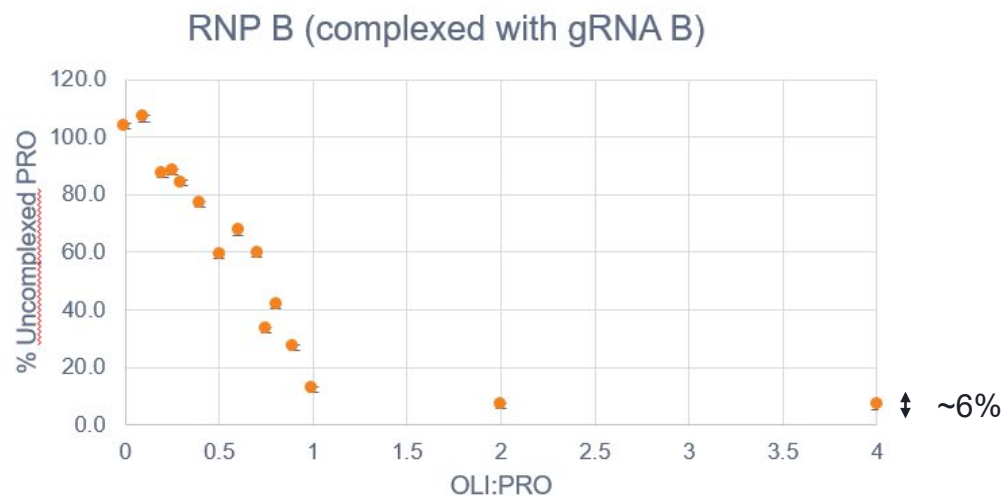
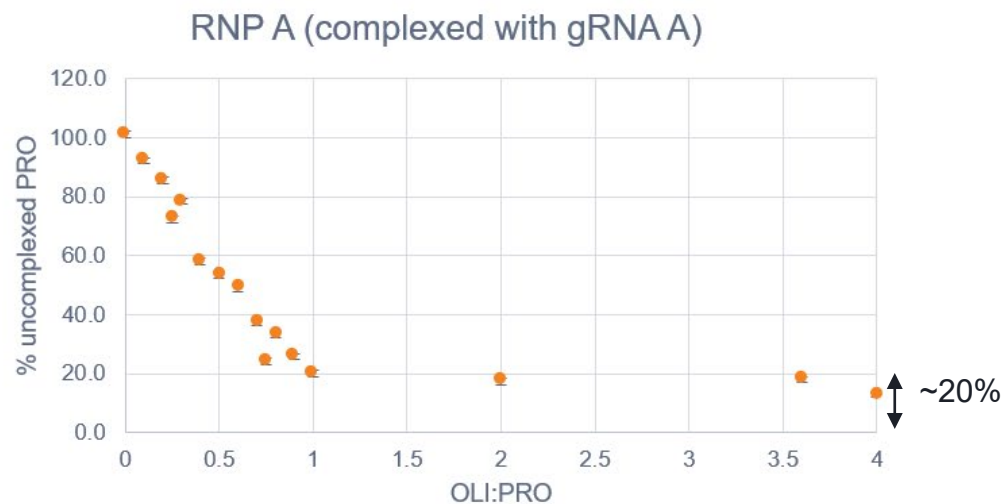
See Poster T.13 (Lemerancier et al.)







Accuracy via WCX-WAX (% Uncomplexed PRO vs OLI:PRO Ratio)





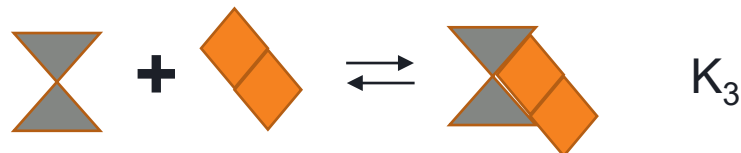
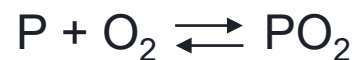
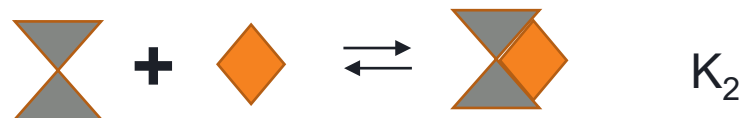
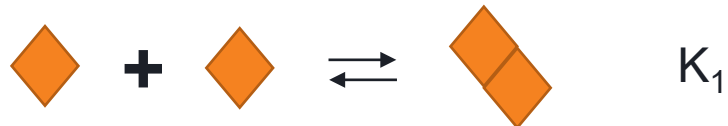
% Uncomplexed PRO vs OLI:PRO Ratio



Different Plateaus for gRNA Monomer vs. Dimer/Multimer?

Possible Explanations:

- kinetically driven
 - *on-column dissociation* $k_{-3} > k_{-2}$
- thermodynamically driven
 - e.g., $K_1, K_2 \gg K_3$
- *modeling not adequate to reproduce data*

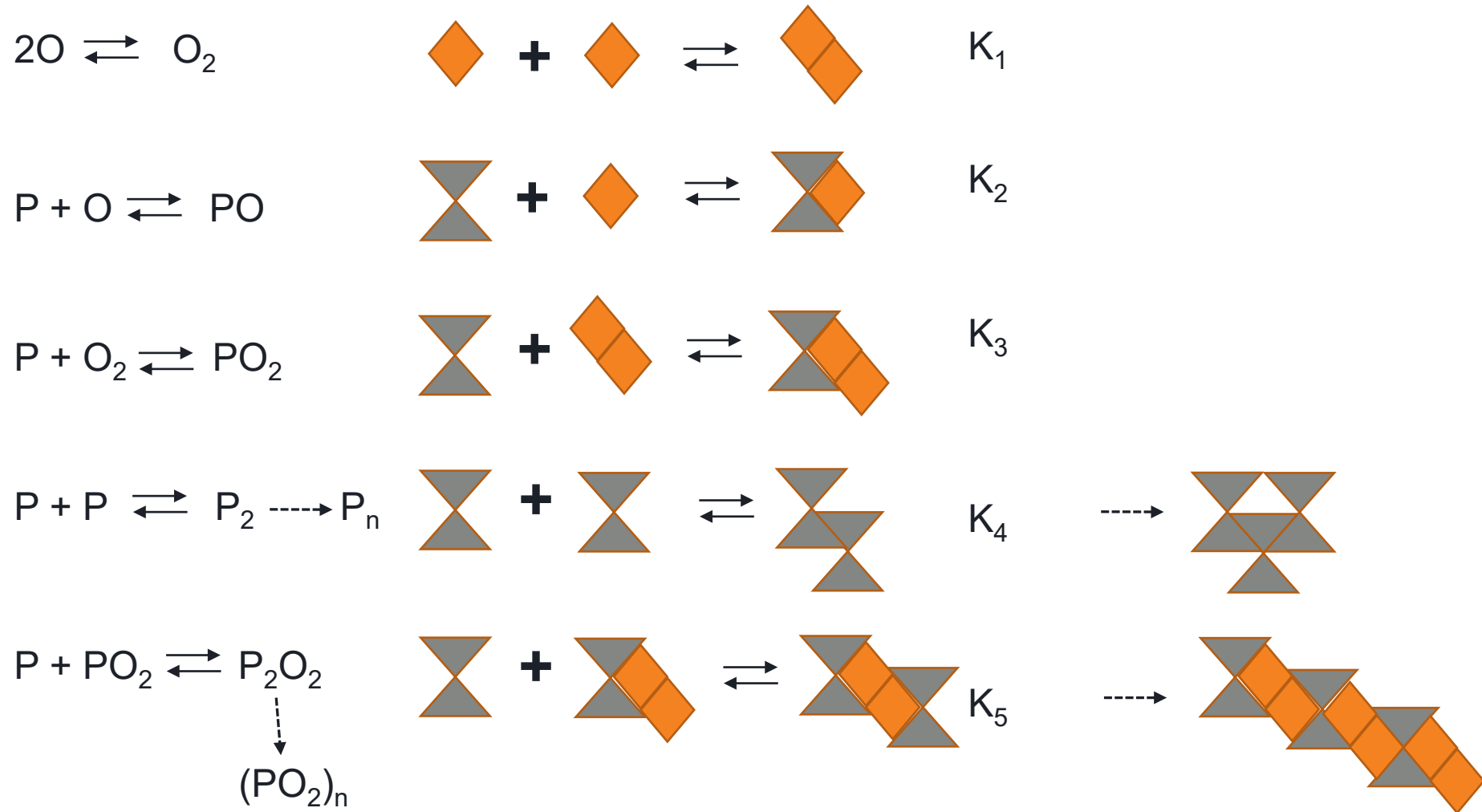




% Uncomplexed PRO vs OLI:PRO Ratio



It's probably a bit more complicated... (analysis in progress)





Analytical Characterization of gRNAs and RNPs

- gRNA Conformers and Multimers
 - formation is sequence dependent, and more likely for longer sequences
 - manifestation is condition dependent (gRNA conformers, stoichiometries, dynamics, and analysis method)
- RNPs
 - another level of complexity due to noncovalent nature of complex (truncates etc., conformers, stoichiometries, dynamics, and the analysis method)
 - SEC, IEX, PAGE, IM data shows that the secondary structures formed by the guides impact the complexation
 - some method dependence of results, which is not surprising for these noncovalent complexes of large biomolecules
 - “Combo” WCX/WAX method is promising as a characterization and possible release method



Thank You